*pathogens*

MDPI

*Article*

# Upscaling the surveillance of tick-borne pathogens in the French Caribbean islands

**Mathilde Gondard [1,2], Sabine Delannoy [3], Valérie Pinarello [2,4], Rosalie Aprelon [2,4], Elodie Devillers [1], Clémence Galon [1], Jennifer Pradel [2,4], Muriel Vayssier-Taussat [1], Emmanuel Albina [2,4] and Sara Moutailler [1*]**

[1] UMR BIPAR, Animal Health Laboratory, ANSES, INRA, Ecole Nationale Vétérinaire d'Alfort, Université Paris-Est, Maisons-Alfort, France; mathilde.gondard@gmail.com; elodie.devilliers@anses.fr; clemence.galon@anses.fr; Muriel.Vayssier@inra.fr; sara.moutailler@anses.fr

[2] CIRAD, UMR ASTRE, F-97170 Petit-Bourg, Guadeloupe, France; mathilde.gondard@gmail.com; valerie.pinarello@cirad.fr; rosalie.aprelon@cirad.fr; jennifer.pradel@cirad.fr; emmanuel.albina@cirad.fr

[3] IdentyPath Platform, Laboratory for Food Safety, ANSES, Maisons-Alfort, France. Sabine.delannoy@anses.fr

[4] ASTRE, Univ Montpellier, CIRAD, INRA, Montpellier, France; valerie.pinarello@cirad.fr; rosalie.aprelon@cirad.fr; jennifer.pradel@cirad.fr; emmanuel.albina@cirad.fr

* Correspondence: sara.moutailler@anses.fr; Tel.: +33 1 49 77 46 50

Received: date; Accepted: date; Published: date

Please note that this paper has already been published online as a preprint on bioRxiv: https://doi.org/10.1101/532457

**Abstract:** Despite the high burden of vector-borne disease in (sub)-tropical areas, few information are available regarding the diversity of tick and tick-borne pathogens circulating in the Caribbean. Management and control of vector-borne disease require actual epidemiological data to better assess and anticipate the risk of (re)-emergence of tick-borne diseases in the region. To simplify and reduce the costs of such large-scale surveys, we implemented a high-throughput microfluidic real-time PCR system suitable for the screening of the main bacterial and parasitic genera involved in tick-borne disease and potentially circulating in the area. We used the new screening tool to perform an exploratory epidemiological study on 132 specimens of *Amblyomma variegatum* and 446 of *Rhipicephalus microplus* collected in Guadeloupe and Martinique. Not only the system was able to detect the main pathogens of the area– *Ehrlichia ruminantium*, *Rickettsia africae*, *Anaplasma marginale*, *Babesia bigemina*, *Babesia bovis*– but the system also provided evidence of unsuspected microorganisms in Caribbean ticks, belonging to the *Anaplasma*, *Ehrlichia*, *Borrelia* and *Leishmania* genera. Our study demonstrated how high-throughput microfluidic real-time PCR technology can assist large-scale epidemiological studies, providing a rapid overview of tick-borne pathogen and microorganism diversity, and opening up new research perspectives for the epidemiology of tick-borne pathogens.

**Keywords:** Tick; Bacteria; Parasites; Caribbean; Microfluidic real-time PCRs.

## 1. Introduction

Among hematophagous arthropods, ticks transmit the greatest variety of pathogens of public health and veterinary importance whose incidence is growing worldwide [1]. The French West Indies, including the islands of Guadeloupe and Martinique, are located in the heart of the Caribbean Neotropical zone, a cosmopolitan area characterized by a tropical climate, intercontinental trade and animal movements (legal and illegal trade as well as bird migration) that are favorable for the introduction and spread of ticks and tick-borne pathogens (TBPs) [2]. Yet, the epidemiological

situation of the Caribbean area with regard to the diversity of tick species and tick-borne diseases (TBDs) is poorly documented [3].

*Amblyomma variegatum*, also known as the tropical bont tick (TBT) in the Caribbean, and *Rhipicephalus microplus* (the "cattle tick") have been the two main tropical livestock pests since their introduction in the Caribbean through imports of infested animals from Africa and Asia in the 18th-19th centuries [4]. Both tick species are present in the French West Indies, where they are involved in the transmission of TBPs of medical and veterinary importance [5-9].

*R. microplus*, a one-host tick highly specific to cattle, is mainly involved in the transmission of *Anaplasma marginale*, *Babesia bovis* and *Babesia bigemina*, causing bovine anaplasmosis and babesiosis, respectively. These endemic pathogens are responsible for important economical lost to farming industries in the Caribbean and are still a sanitary threat [7,10].

*A. variegatum* is a three-host tick species, with immature stages that can parasitize a wide range of hosts, including rodents, mongooses and birds, as well as an adult stage that is more specific to cattle [11]. This tick species is mainly involved in *Ehrlichia ruminantium* transmission, the causative agent of heartwater, a fatal ruminant ehrlichiosis. Although *A. variegatum* is present in both Martinique (mainly in the south) and Guadeloupe (widespread), *E. ruminantium* has only been reported in Guadeloupe [12]. In addition, *A. variegatum* ticks are also a vector of *Rickettsia africae*, which is common in the Caribbean and can induce human rickettsiosis, called African tick-bite fever [9,13,14]. African tick-bite fever remains a concern mainly for travelers. Indeed, despite high levels of tick infection and seroprevalence in human and cattle sera, only two human cases of African tick-bite fever have been reported to date, only in travelers returning from Guadeloupe [9,15]. Lastly, *A. variegatum* is also involved in the epidemiology of *Theileria mutans* and *Theileria velifera*, two cattle parasites with low and no virulence, respectively [6,8]. However, very few information are available on the distribution and prevalence of these two apicomplexa in the Carribean.

Most of the epidemiological data available did not survey or determine the diversity of TBPs circulating in the Carribean, since they were often limited to the detection of some well-known pathogens, via serological studies in animals or humans, or on molecular biology testing (PCR, nested PCR) [16,17]. Thus, regarding the lack of recent information, the limited extent of the epidemiological data available, new insight into the epidemiology of ticks and TBPs was needed to better address the prevalence and (re-)emergence of TBDs in the Caribbean.

In order to improve the surveillance ability of tick-borne pathogens in the Neotropical area, we implemented a new large scale screening tool based on microfluidic real-time PCR approach. Microfluidic real-time PCR system is based on the use of microfluidic chip allowing the performance of up to 9216 individual PCR reactions per run, and thus the simultaneous detection of up to 96 target into up to 96 samples. The recent development and use of a microfluidic real-time PCR system for the rapid and concomitant detection of a large panel of TBPs in European ticks has paved the way for promising and broader surveillance capacities [18-22]. Here, we adapted and designed a new microfluidic real-time PCR system suited to the simultaneous screening of the main bacteria and protozoans potentially transmitted by ticks in the Caribbean. Not only did the system enable the direct detection of 49 bacterial and parasitic species, but it also enabled, within a single experiment, broader capacities for the surveillance of potentially pathogenic microorganisms, by targeting the main bacterial and protozoan genera involved in human and animal vector-borne diseases (one protozoan phylum and eight bacterial and protozoan genera). In addition, the system enabled the molecular identification of the three well-known tick species involved in TBDs in the Caribbean in order to confirm the morphological tick species identification determined on the field. Finally, we used the new high-throughput detection tool to conduct large-scale screening of TBPs in 132 *A. variegatum* and 446 *R. microplus* specimens collected in Guadeloupe and Martinique. We demonstrated the system's ability to detect well-known TBPs occurring in the French West Indies, as well as unsuspected TBPs and potential new microorganisms. This new method can considerably improve the ability to monitor emerging and non-emerging TBPs through large-scale surveys in the Caribbean area.

## 2. Results

### 2.1 *Implementation of the high-throughput microfluidic real-time PCR system for tick-borne pathogen screening*

The high-throughput microfluidic real-time PCR system developed for the screening of known and potential TBPs in Caribbean ticks included 61 sets of primers and probes. Among them, 49 designs were developed for the detection of bacterial (n=32) and protozoan (n=17) species and bacterial (n=5) and protozoan (n=3) genera/phyla (Table 1). Three sets of primers and probes were developed for the molecular identification of the three tick species found in the Caribbean: *A. variegatum*, *R. microplus* and *R. sanguineus* sensu lato (Table 1). Lastly, we developed a design targeting a conserved region of the 16S rRNA genes in ticks, called "Tick spp.", used as a control for DNA/RNA extraction (Table 1).

**Table 1.** List of primer/probe sets constituting the BioMark system, with the positive controls used for their validation.

| Microorganisms | Target | Design name | Sequence (5′ à 3′) | Length (bp) | Positive controls |
|---|---|---|---|---|---|
| *Rickettsia* spp. | gltA | Rick_spp_gltA_F | GTCGCAAATGTTCACGGTACTT | 78 | **, Culture of *R. slovaca* |
| | | Rick_spp_gltA_R | TCTTCGTGCATTTCTTTCCATTG | | |
| | | Rick_spp_gltA_P | TGCAATAGCAAGAACCGTAGGCTGGATG | | |
| *Rickettsia massiliae* * | ITS | Ri_ma_ITS_F | GTTATTGCATCACTAATGTTATACTG | 128 | Culture |
| | | Ri_ma_ITS_R | GTTAATGTTGTTGCACGACTCAA | | |
| | | Ri_ma_ITS_P | TAGCCCCGCCACGATATCTAGCAAAAA | | |
| *Rickettsia rickettsii* * | ITS | Ri_ri_ITS_F | TCTACTCACAAAGTTATCAGGTTAA | 124 | Plasmid |
| | | Ri_ri_ITS_R | CCTACGATACTCAGCAAAATAATTT | | |
| | | Ri_ri_ITS_P | TCGCTGGATATCGTTGCAGGACTACAG | | |
| *Rickettsia conorii* | sca1 | Ri_co_sca1_F | GTAGATGCTTCATAGAATACTGC | 88 | Infected *Rhipicephalus sanguineus* s.l. |
| | | Ri_co_sca1_R | CCAAATTTAGTCTACCTTGTGATC | | |
| | | Ri_co_sca1_P | TCCTCCTGACGTATTAAAAGAAGCTGAAGCT | | |
| *Rickettsia africae* | sca1 | Ri_af_sca1_F | GATACGACAAGTACCTCGCAG | 122 | Culture |
| | | Ri_af_sca1_R | GGATTATATACTTTAGGTTCGTTAG | | |
| | | Ri_af_sca1_P | CAGATAGGAACAGTAATTGTAACGGAACCAG | | |
| *Rickettsia felis* | orfB | Ri_fel_orfB_F | ACCCTTTTCGTAACGCTTTGC | 163 | Culture |
| | | Ri_fel_orfB_R | TATACTTAATGCTGGGCTAAACC | | |
| | | Ri_fel_orfB_P | AGGGAAACCTGGACTCCATATTCAAAAGAG | | |
| *Rickettsia typhi* | ompB | Ri_typ_ompB_F | CAGGTCATGGTATTACTGCTCA | 133 | Culture |
| | | Ri_typ_ompB_R | GCAGCAGTAAAGTCTATTGATCC | | |
| | | Ri_typ_ompB_P | ACAAGCTGCTACTACAAAAAGTGCTCAAAATG | | |
| *Rickettsia prowazekii* | gltA | Ri_pro_gltA_F | CAAGTATCGGTAAAGATGTAATCG | 151 | Plasmid |
| | | Ri_pro_gltA_R | TATCCTCGATACCATAATATGCC | | |
| | | Ri_pro_gltA_P | ATATAAGTAGGGTATCTGCGGAAGCCGAT | | |

| | | | | | |
|---|---|---|---|---|---|
| *Borrelia* spp. * | 23S rRNA | Bo_bu_sl_23S_F | GAGTCTTAAAAGGGCGATTTAGT | 73 | **, Culture of *B. afzelii*, *B. garinii*, *B. valaisiana*, *B. lusitaniae*, *B. recurrentis* |
| | | Bo_bu_sl-23S_R | CTTCAGCCTGGCCATAAATAG | | |
| | | Bo_bu_sl_23S_P | TAGATGTGGTAGACCCGAAGCCGAGT | | |
| *Borrelia burgdorferi* sensu stricto | glpA | Bo_bu_glpA_F | GCAATTACAAGGGGGTATAAAGC | 206 | Culture |
| | | Bo_bu_glpA_R | GGCGTGATAAGTGCACATTCG | | |
| | | Bo_bu_glpA_P | TTAATTAAACGGGGTGCATTCTTCTCAAGAATG | | |
| *Borrelia anserina* | fla | Bor_ans_fla_F | GGAGCACAACAAGAGGGAG | 76 | Plasmid |
| | | Bor_ans_fla_R | TTGGAGAATTAACCCCACCTG | | |
| | | Bor_ans_fla_P | TGCAAGCAACTCCAGCTCCAGTAGCT | | |
| *Borrelia lonestari* | glpQ | Bor_lon_glpQ_F | GATCCAGAACTTGATACAACCAC | 99 | Infected *Amblyomma americanum* |
| | | Bor_lon_glpQ_R | TTCATCTAGTGAGAAGTCAGTAG | | |
| | | Bor_lon_glpQ_P | AGTAATATCGTCCGTCTTCCCTAGCTCG | | |
| *Borrelia parkeri* | gyrB | Bor_par_gyrB_F | GCAAAACGATTCAAAGTGAGTCC | 184 | Culture |
| | | Bor_par_gyrB_R | CTCATTGCCTTTAAGAAACCACTT | | |
| | | Bor_par_gyrB_P | TTAAAACCAGCAACATGAGTTCCTCCTTCTC | | |
| *Borrelia bissettii* * | rpoB | Bo_bi_rpoB_F | GCAACCAGTCAGCTTTCACAG | 118 | Plasmid |
| | | Bo_bi_rpoB_R | CAAATCCTGCCCTATCCCTTG | | |
| | | Bo_bi_rpoB_P | AAAGTCCTCCCGGCCCAAGAGCATTAA | | |
| *Borrelia theileri* | glpQ | Bo_th_glpQ_F | GTGCTAACAAAGGACAATATTCC | 213 | Plasmid |
| | | Bo_th_glpQ_R | GGTTAGTGGAAAACGGTTAGGAT | | |
| | | Bo_th_glpQ_P | TATTATAATTCACGAGCCAGAGCTTGACAC | | |
| *Bartonella* spp. | ssrA | Bart_spp_ssrA_F | CGTTATCGGGCTAAATGAGTAG | 118 | **, Culture of *B. quintana* |
| | | Bart_spp_ssrA_R | ACCCCGCTTAAACCTGCGA | | |
| | | Bart_spp_ssrA_P | TTGCAAATGACAACTATGCGGAAGCACGTC | | |
| *Bartonella barcilliformis* * | rpoB | Ba_ba_rpoB_F | GAAGAGTTTGTAGTTTGTCGTCA | 105 | Culture |
| | | Ba_ba_rpoB_R | AGCAGCTACAGAAACCAACTG | | |
| | | Ba_ba_rpoB_P | TGCAGGTGAAGTTTTGATGGTGCCACG | | |
| *Bartonella henselae* | ribC | Bar_he_ribC_F | GGGATGCGATTTAATAGTTCTAC | 116 | Culture |
| | | Bar_he_ribC_R | CGCTTGTTGTTTTGATCCTCG | | |
| | | Bar_he_ribC_P | ACGTTATAGTAGCGAAAACTTAGAAATTGGTGC | | |
| *Bartonella vinsonii subsp. berkhoffii* | ITS | Bar_vin_ITS_F | GGAATTGCTTAACCCACTGTTG | 141 | Culture |
| | | Bar_vin_ITS_R | CCTTATTGATTTAGATCTGATGGG | | |
| | | Bar_vin_ITS_P2 | AGAAACTCCCGCCTTTATGAGAGAAATCTCT | | |
| | Icd | Co_bu_icd_F | AGGCCCGTCCGTTATTTTACG | 74 | Culture |
| | | Co_bu_icd_R | CGGAAAATCACCATATTCACCTT | | |

| | | | | | |
|---|---|---|---|---|---|
| | | Co_bu_icd_P | TTCAGGCGTTTTGACCGGGCTTGGC | | |
| *Coxiella burnetii and Coxiella-like* * | IS1111 | Co_bu_IS111_F | TGGAGGAGCGAACCATTGGT | 86 | Culture |
| | | Co_bu_IS111_R | CATACGGTTTGACGTGCTGC | | |
| | | Co_bu_IS111_P | ATCGGACGTTTATGGGGATGGGTATCC | | |
| | | Fr_tu_tul4_F | ACCCACAAGGAAGTGTAAGATTA | | |
| *Francisella tularensis and Francisella-like endosymbionts* * | tul4 | Fr_tu_tul4_R | GTAATTGGGAAGCTTGTATCATG | 76 | Culture |
| | | Fr_tu_tul4_P | AATGGCAGGCTCCAGAAGGTTCTAAGT | | |
| | | Fr_tu_fopA_F | GGCAAATCTAGCAGGTCAAGC | | |
| | fopA | Fr_tu_fopA_R | CAACACTTGCTTGAACATTTCTAG | 91 | Culture |
| | | Fr_tu_fopA_P | AACAGGTGCTTGGGATGTGGGTGGTG | | |
| *Anaplasma* spp. | 16S rRNA | Ana_spp_16S_F | CTTAGGGTTGTAAAACTCTTTCAG | | |
| | | Ana_spp_16S_R | CTTTAACTTACCAAACCGCCTAC | 160 | ** |
| | | Ana_spp_16S_P | ATGCCCTTTACGCCCAATAATTCCGAACA | | |
| *Anaplasma marginale* * | msp1b | An_ma_msp1_F | CAGGCTTCAAGCGTACAGTG | | Experimentally infected bovine blood sample |
| | | An_ma_msp1_R | GATATCTGTGCCTGGCCTTC | 85 | |
| | | An_ma_msp1_P | ATGAAAGCCTGGAGATGTTAGACCGAG | | |
| *Anaplasma phagocytophilum* * | msp2 | An_ph_msp2_F | GCTATGGAAGGCAGTGTTGG | | Infected *Ixodes* spp. tick |
| | | An_ph_msp2_R | GTCTTGAAGCGCTCGTAACC | 77 | |
| | | An_ph_msp2_P | AATCTCAAGCTCAACCCTGGCACCAC | | |
| *Anaplasma platys* * | groEL | An_pla_groEL_F | TTCTGCCGATCCTTGAAAACG | | Infected canine blood sample |
| | | An_pla_groEL_R | CTTCTCCTTCTACATCCTCAG | 75 | |
| | | An_pla_groEL_P | TTGCTAGATCCGGCAGGCCTCTGC | | |
| *Anaplasma bovis* * | groEL | An_bo_groEL_F | GGGAGATAGTACACATCCTTG | | Plasmid |
| | | An_bo_groEL_R | CTGATAGCTACAGTTAAGCCC | 73 | |
| | | An_bo_groEL_P | AGGTGCTGTTGGATGTACTGCTGGACC | | |
| *Anaplasma ovis* * | msp4 | An_ov_msp4_F | TCATTCGACATGCGTGAGTCA | | Plasmid |
| | | An_ov_msp4_r | TTTGCTGGCGCACTCACATC | 92 | |
| | | An_ov_msp4_P | AGCAGAGAGACCTCGTATGTTAGAGGC | | |
| *Ehrlichia* spp. * | 16S rRNA | Neo_mik_16S_F | GCAACGCGAAAAACCTTACCA | | |
| | | Neo_mik_16S_R | AGCCATGCAGCACCTGTGT | 98 | ** |
| | | Neo_mik_16S_P | AAGGTCCAGCCAAACTGACTCTTCCG | | |
| | gltA | Eh_ca_gltA_F | GACCAAGCAGTTGATAAAGATGG | 136 | Culture |

| | | | | | |
|---|---|---|---|---|---|
| *Ehrlichia canis* | dsb | Eh_ca_gltA_R | CACTATAAGACAATCCATGATTAGG | 117 | Infected *Amblyomma americanum* |
| | | Eh_ca_gltA_P | ATTAAAACATCCTAAGATAGCAGTG GCTAAGG | | |
| *Ehrlichia chaffeensis* * | | Eh_ch_dsb_F | TATTGCTAATTACCCTCAAAAAGTC | | |
| | | Eh_ch_dsb_R | GAGCTATCCTCAAGTTCAGATTT | | |
| | | Eh_ch_dsb_P | ATTGACCTCCTAACTAGAGGGCAAG CA | | |
| *Ehrlichia ewingii* * | dsb | Eh_ew_dsb_F | CAATACTTGGAGAAGCATCATTG | 111 | Infected *Amblyomma americanum* |
| | | Eh_ew_dsb_R | TTGCTTATGGCTTAATGCTGCAT | | |
| | | Eh_ew_dsb_P | AAAGCAGTACGTGCAGCATTGGCTG TA | | |
| *Ehrlichia ruminantium* | gltA | Eh_ru_gltA_F | CCAGAAAACTGATGGTGAGTTAG | 116 | Culture |
| | | Eh_ru_gltA_R | AGCCTACATCAGCTTGAATGAAG | | |
| | | Eh_ru_gltA_P | AGTGTAAACTTGCTGTTGCTAAGGTA GCATG | | |
| *Panola Mountain Ehrlichia* | gltA | Eh_PME_gltA_F | GCTAGTTATGAGTTAGAATGTAAAC | 121 | Infected *Amblyomma americanum* |
| | | Eh_PME_gltA_R | TACTATAGGATAATCTTGAATCAGC | | |
| | | Eh_PME_gltA_P | TTGCTATCGCTAAAATTCCAAGTATG ATTGCG | | |
| *Neoehrlichia mikurensis* * | groEL | Neo_mik_groEL_F | AGAGACATCATTCGCATTTTGGA | 96 | Infected rodent blood sample |
| | | Neo_mik_groEL_R | TTCCGGTGTACCATAAGGCTT | | |
| | | Neo_mik_groEL_P | AGATGCTGTTGGATGTACTGCTGGAC C | | |
| *Aegyptianella pullorum* | groEL | Ae_pul_groEL_F | AGCCAGTATTATCGCTCAAGG | 168 | Plasmid |
| | | Ae_pul_groEL_R | GCCTCACGTGCCTTCATAAC | | |
| | | Ae_pul_groEL_P | TGCTTCTCAGTGTAACGACAGGGTTG G | | |
| *Apicomplexa* | 18S rRNA | Apic_18S_F | TGAACGAGGAATGCCTAGTATG | 104 | **, Infected canine blood sample, with *B. canis rossi*, *B. canis canis*; Culture of *B. divergens*, *T. lestoquari*, *T. annulata* |
| | | Apic_18S_R | CACCGGATCACTCGATCGG | | |
| | | Apic_18S_S | TAGGAGCGACGGGCGGTGTGTAC | | |
| *Babesia canis vogeli* * | hsp70 | Ba_vo_hsp70_F | TCACTGTGCCTGCGTACTTC | 87 | Infected canine blood sample |
| | | Ba_vo_hsp70_R | TGATACGCATGACGTTGAGAC | | |
| | | Ba_vo_hsp70_P | AACGACTCCCAGCGCCAGGCCAC | | |
| *Babesia ovis* * | 18S rRNA | Ba_ov_18S_F | TCTGTGATGCCCTTAGATGTC | 92 | Plasmid |
| | | Ba_ov_18S_R | GCTGGTTACCCGCGCCTT | | |
| | | Ba_ov_18S_P | TCGGAGCGGGGTCAACTCGATGCAT | | |
| *Babesia bigemina* * | | Ba_big_RNA18S_F | ATTCCGTTAACGAACGAGACC | 99 | Plasmid |

| | | | | | |
|---|---|---|---|---|---|
| | 18S rRNA | Ba_big_RNA18S_R | TTCCCCCACGCTTGAAGCA | | |
| | | Ba_big_RNA18S_P | CAGGAGTCCCTCTAAGAAGCAAACGAG | | |
| *Babesia gibsoni* | Rap1 | Ba_gib_rap1_F | CTCTTGCTCATCATCTTTTCGG | 130 | Plasmid |
| | | Ba_gib_rap1_R | TCAGCGTATCCATCCATTATATG | | |
| | | Ba_gib_rap1_S | TTTAATGCGTGCTACGTTGTACTTCCCAAAG | | |
| *Babesia caballi* * | Rap1 | Ba_cab_rap1_F | GTTGTTCGGCTGGGGCATC | 94 | Plasmid |
| | | Ba_cab_rap1_R | CAGGCGACTGACGCTGTGT | | |
| | | Ba_cab_rap1_P | TCTGTCCCGATGTCAAGGGGCAGGT | | |
| *Babesia bovis* * | CCTeta | Ba_bo_CCTeta_F | GCCAAGTAGTGGTAGACTGTA | 100 | Plasmid |
| | | Ba_bo_CCTeta_R | GCTCCGTCATTGGTTATGGTA | | |
| | | Ba_bo_CCTeta_P | TAAAGACAACACTGGGTCCGCGTGG | | |
| *Babesia duncani* * | ITS2 | Ba_du_ITS_F | ATTTCCGTTTGCGAGAGTTGC | 87 | Plasmid |
| | | Ba_du_ITS_R | AGGAAGCATCAAGTCATAACAAC | | |
| | | Ba_du_ITS_P | AACAAGAGGCCCCGAGATCAAGGCAA | | |
| *Babesia microti* * | CCTeta | Bab_mi_CCTeta_F | ACAATGGATTTTCCCCAGCAAAA | 145 | Culture |
| | | Bab_mi_CCTeta_R | GCGACATTTCGGCAACTTATATA | | |
| | | Bab_mi_CCTeta_P | TACTCTGGTGCAATGAGCGTATGGGTA | | |
| *Theileria parva* * | 18S rRNA | Th_pa_18S_F | GAGTATCAATTGGAGGGCAAG | 173 | Culture |
| | | Th_pa_18S_R | CAGACAAAGCGAACTCCGTC | | |
| | | Th_pa_18S_P | AAATAAGCCACATGCAGAGACCCCGAA | | |
| *Theileria mutans* | ITS | The_mu_ITS_F | CCTTATTAGGGGCTACCGTG | 119 | Plasmid |
| | | The_mu_ITS_R | GTTTCAAATTTGAAGTAACCAAGTG | | |
| | | The_mu_ITS_P | ATCCGTGAAAAACGTGCCAAACTGGTTAC | | |
| *Theileria velifera* | 18S rRNA | The_ve_18S_F | TGTGGCTTATCTGGGTTCGC | 151 | Plasmid |
| | | The_ve_18S_R | CCATTACTTTGGTACCTAAAACC | | |
| | | The_ve_18S_P | TTGCGTTCCCGGTGTTTTACTTTGAGAAAG | | |
| *Theileria equi* | ema1 | Th_eq_ema1_F4 | CGGCAAGAAGCACACCTTC | 167 | Plasmid |
| | | Th_eq_ema1_R4 | TGCCATCGCCCTTGTAGAG | | |
| | | Th_eq_ema1_P4 | AAGGCTCCAGGCAAGCGCGTCCT | | |
| *Cytauxzoon felis* | ITS2 | Cy_fel_ITS2_F | AAGATCCGAACGGAGTGAGG | 119 | Plasmid |
| | | Cy_fel_ITS2_R | GTAGTCTCACCCAATTTCAGG | | |
| | | Cy_fel_ITS2_S | AAGTGTGGGATGTACCGACGTGTGAG | | |
| *Hepatozoon* spp. | 18S rRNA | Hepa_spp_18S_F | ATTGGCTTACCGTGGCAGTG | 175 | ** |
| | | Hepa_spp_18S_R | AAAGCATTTTAACTGCCTTGTATTG | | |
| | | Hepa_spp_18S_S | ACGGTTAACGGGGGATTAGGGTTCGAT | | |

| | | | | | |
|---|---|---|---|---|---|
| *Hepatozoon canis* | 18S rRNA | He_can_18S_F | TTCTAACAGTTTGAGAGAGGTAG | 221 | Infected canine blood sample |
| | | He_can_18S_R | AGCAGACCGGTTACTTTTAGC | | |
| | | He_can_18S_S | AGAACTTCAACTACGAGCTTTTTAACTGCAAC | | |
| *Hepatozoon americanum* | 18S rRNA | He_ame_18S_F2 | GGTATCATTTTGGTGTGTTTTTAAC | 159 | Plasmid |
| | | He_ame_18S_R2 | CTTATTATTCCATGCTCCAGTATTC | | |
| | | He_ame_18S_P2 | AAAAGCGTAAAAGCCTGCTAAAAACACTCTAC | | |
| *Leishmania* spp. | hsp70 | Leish_spp_hsp70_F | CGACCTGTTCCGCAGCAC | 78 | ** and culture of *L. martiniquensis* |
| | | Leish_spp_hsp70_R | TCGTGCACGGAGCGCTTG | | |
| | | Leish_spp_hsp70_S | TCCATCTTCGCGTCCTGCAGCACG | | |
| *Leishmania infantum* | ITS | Le_inf_ITS_F | CGCACCGCCTATACAAAAGC | 103 | Culture |
| | | Le_inf_ITS_R | GTTATGTGAGCCGTTATCCAC | | |
| | | Le_inf_ITS_S | ACACGCACCCACCCCGCCAAAAAC | | |
| *Rangelia vitalii* | 18S rRNA | Ra_vit_18S_F | TAACCGTGCTAATTGTAGGGC | 92 | Plasmid |
| | | Ra_vit_18S_R | GAATCACCAAACCAAATGGAGG | | |
| | | Ra_vit_18S_S | TAATACACGTTCGAGGGCGCGTTTTGC | | |
| *Tick* spp. | 16S rRNA | Tick_spp_16S_F | AAATACTCTAGGGATAACAGCGT | 99 | ** |
| | | Tick_spp_16S_R | TCTTCATCAAACAAGTATCCTAATC | | |
| | | Tick_spp_16S_P | CAACATCGAGGTCGCAAACCATTTTGTCTA | | |
| *Amblyomma variegatum* | ITS2 | Amb_var_ITS2_F | GCCAGCCTCTGAAGTGACG | 117 | Tick extract (Guadeloupe) |
| | | Amb_var_ITS2_R | TTCTGCGGTTTAAGCGACGC | | |
| | | Amb_var_ITS2_P | TCTTGCCACTCGACCCGTGCCTC | | |
| *Rhipicephalus microplus* | ITS2 | Rhi_mic_ITS2_F | GCTTAAGGCGTTCTCGTCG | 144 | Tick extract (Galapagos Islands) |
| | | Rhi_mic_ITS2_R | CAAGGGCAGCCACGCAG | | |
| | | Rhi_mic_ITS2_P | TAGTCCGCCGTCGGTCTAAGTGCTTC | | |
| *Rhipicephalus sanguineus* sensu lato | ITS2 | Rhi_san_ITS2_F | TTGAACGCTACGGCAAAGCG | 110 | Tick extract (France) |
| | | Rhi_san_ITS2_R | CCATCACCTCGGTGCAGTC | | |
| | | Rhi_san_ITS2_P | ACAAGGGCCGCTCGAAAGGCGAGA | | |

The detection ability of each design and the effect of pre-amplification on detection signals were first checked by TaqMan real-time PCR on a LightCycler 480 apparatus using a range of dilutions of positive controls (Table 1, Table S2). Three kinds of positive controls were used, including bacterial or protozoan cultures when available, DNA from infected ticks or blood samples, and plasmidic constructions as a last resort (Table 1). Except for the design targeting *Borrelia burgdorferi* sensu stricto, which never succeeded in detecting the positive controls even after a pre-amplification step, the remaining 60 designs targeting TBPs and tick species were able to detect their target with Ct values

between six and 38 (data not shown). Pre-amplification improved the quality of detection and was therefore validated as part of the screening protocol (see Figure S1).

The relative specificity of the 61 designs was then evaluated using the BioMark system and a total of 62 positive controls (Figure 1, Table S2).

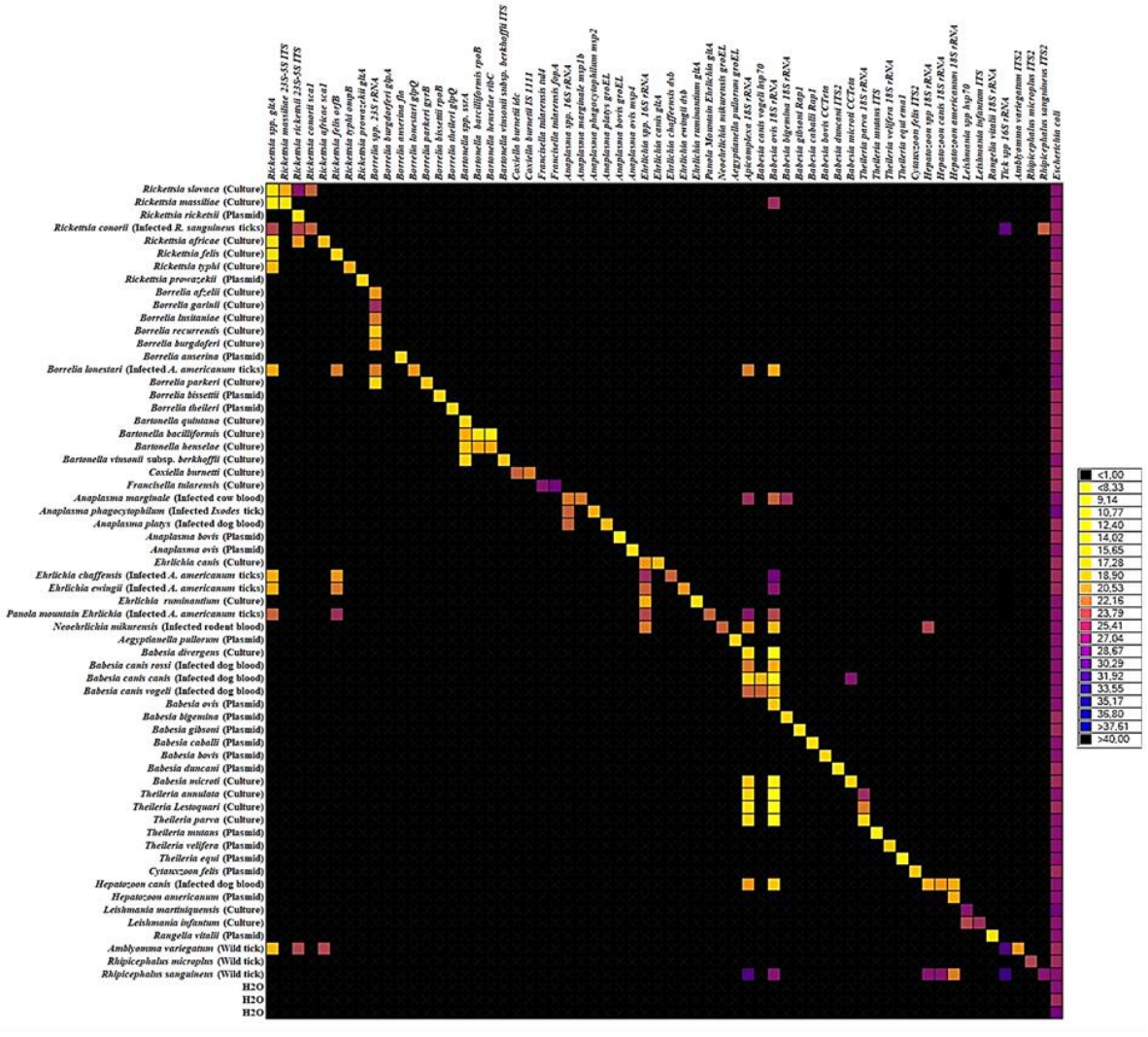



**Figure 1.** BioMarkTM dynamic array system specificity test (96.96 chip). Each square corresponds to a single real-time PCR reaction, where rows indicate the pathogen in the sample and columns represent the target of the primer/probe set. Ct values for each reaction are represented by a color gradient; the color scale is shown on the right y-axis. The darkest shades of blue and black squares are considered as negative reactions with Ct > 30.

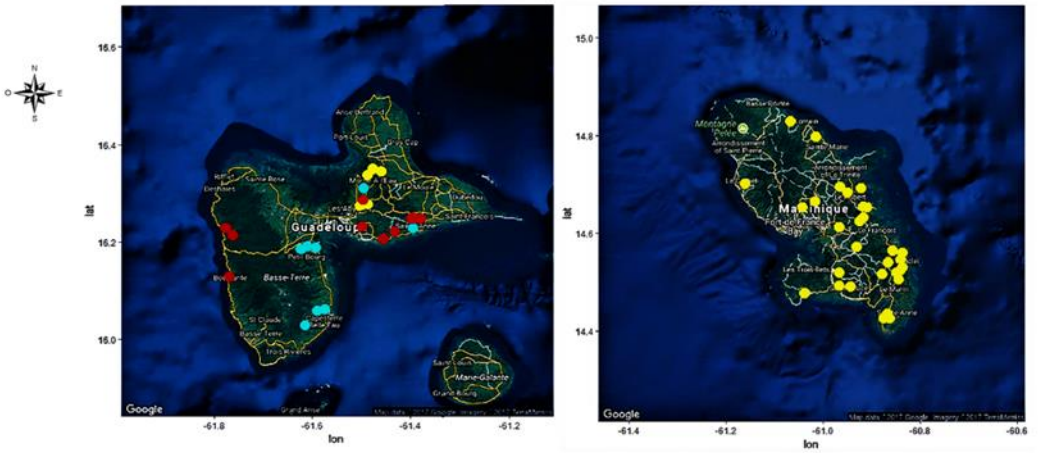
Forty-three primer/probe sets were able to specifically detect and amplify their target using a Ct cut-off value of 30; they were then directly validated (Figure 1). The remaining designs were able to detect and amplify their target, but they also gave positive results in outgroup controls. Interestingly, two kinds of unsuspected signals were observed; some were related to cross-reactions with closely related species and some to potential co-infections in controls corresponding to field samples (Figure 1). Thus, eight designs – *Rickettsia massiliae*, *Rickettsia conorii*, *Bartonella henselae*, *Bartonella bacilliformis*, *Babesia canis vogeli*, *Babesia microti*, *Theileria parva*, *Hepatozoon americanum* – gave positive results in outgroup controls, revealing cross-reactions with one to two closely related species (Figure 1). Caution will be required when interpreting results obtained with these designs. Seven designs – *Rickettsia* spp., *Rickettsia felis*, *Rickettsia africae*, *Apicomplexa*, *Babesia bigemina*, *Hepatozoon* spp., *Hepatozoon canis* – gave positive results in outgroup controls linked to potential co-infection in controls corresponding to DNA from infected ticks or blood samples (Figure 1). As co-infections may

occur in natural tick or blood samples, these unexpected detections in biological samples were likely due to the natural (co-)occurrence of microorganisms rather than to cross-reactions. Finally, the *Babesia ovis* and *Rickettsia rickettsii* designs gave multiple cross-reactions with closely related species or distant outgroups and thus were considered as non-specific and removed from the rest of the study (Figure 1). More details on the relative specificity analysis of the designs are available in Appendix A.

To conclude, with the exception of the sets of primers and probes targeting *Borrelia burgdorferi* sensu stricto, *Babesia ovis* and *Rickettsia rickettsii* that were ultimately removed from the study, the 58 remaining designs were validated for the high-throughput screening of pathogens in Caribbean ticks, taking into account the notified cross-reactions.

### 2.2 Large-scale TBP detection survey in ticks from Guadeloupe and Martinique

A total of 578 adult ticks were collected from cattle in Guadeloupe and Martinique. In total, 523 samples were tested using the BioMarkTM system developed in this study. The Molecular identification of *Amblyomma variegatum* and *Rhipicephalus microplus* using the corresponding specific designs were consistent with the morphological identification made after tick collection. The number of positive ticks and the corresponding infection rates for each detected pathogen were calculated for 132 *A. variegatum* and 165 and 281 *R. microplus* specimens from Guadeloupe and Martinique, respectively (Figure 2). As some of the *R. microplus* samples corresponded to pools of two to four adult specimens, we reported the minimum and maximum infection rates (see Material and methods).




| | Guadeloupe | | Martinique |
|---|---|---|---|
| | Positive *Amblyomma variegatum*, out of 132 (IR) | Positive *Rhipicephalus microplus*, out of 165 (IR min - max) | Positive *Rhipicephalus microplus*, out of 281 (IR min - max) |
| *Anaplasma* spp. | 0 | 8 (4.8%) - 9 (5.5%) | 112 (39.9%) - 116 (41.3%) |
| *Anaplasma marginale* | 0 | 5 (3%) - 7 (4.2%) | 111 (39.5%) - 116 (41.3%) |
| *Ehrlichia* spp. | 11 (8.3%) | 7 (4.2%) - 11(6.7%) | 134 (47.7%) - 138 (49.1%) |
| *Ehrlichia ruminantium* | 7 (5.3%) | 0 | 0 |
| *Borrelia* spp. | 7 (5.3%) | 1 (0.6%) | 12 (4.3%) |
| *Rickettsia* spp. | 130 (98.5%) | 25 (15.2%) - 38 (23%) | 0 |
| *Rickettsia africae* | 126 (95.5%) | 0 | 0 |
| *Leishmania* spp. | 0 | 0 | 2 (0.7%) |
| *Theileria mutans* | 2 (1.5%) | 3 (1.8%) - 4 (2.4%) | 4 (1.4%) |
| *Theileria velifera* | 57 (43.2%) | 39 (23.6%) - 52 (31.5%) | 72 (25.6%) - 73 (26%) |
| *Babesia bigemina* | 0 | 1 (0.6%) - 2 (1.2%) | 35 (12.5%) - 36 (12.8%) |
| *Babesia bovis* | 0 | 0 | 2 (0.7%) |

**Figure 2.** Infection rates in ticks collected in Guadeloupe and Martinique. Number of positive *A. variegatum* ticks (out of 132) and *R. microplus* ticks from Guadeloupe (out of 165) and Martinique (out of 281). Dots on the map indicate the tick collection sites in Guadeloupe and Martinique. Yellow: collection site of *R. microplus* ticks; Red: collection site of *A. variegatum* ticks; Blue: collection site of both tick species, sometimes on the same animal. IR: Infection rate. As some *R. microplus* samples were pooled, we have presented minimum and maximum tick infection rates.

Conventional PCRs/nested PCRs followed by amplicon sequencing were performed on several tick samples to confirm some of the results of the newly designed BioMarkTM system (see Materials and Methods section). Identity percentages of the sequences obtained with reference sequences available in GenBank (NCBI) are presented in Table 2.

**Table 2.** Homology between the deposited sequences and reference sequences in GenBank.

| Biomark Id | Sequence Name | T | S | An | L | Closest Homology | Id% | Reference |
|---|---|---|---|---|---|---|---|---|
| | | 3 | 1 | MK049 | 24 | | | |
| *Rickettsia* spp. | *Rickettsia africae* Tick208 | 0 | 4 | 851 | 8 | *Rickettsia africae* | 100 | AF123706.1 |
| Leishmania | *Leishmania* | | | MK049 | 27 | *Leishmania* | | |
| spp. | *martiniquensis* Tick389 | 2 | 1 | 850 | 2 | *martiniquensis* | 100 | AF303938.1 |
| | | | | | | *Leishmania siamensis* | 100 | GQ226033.1 |
| | | 3 | | MK049 | 24 | | | |
| *Borrelia* spp. | *Borrelia* sp. Tick7 | 0 | 1 | 846 | 5 | *Borrelia anserina* | 90 | X75201.1 |
| | | | | MK049 | 32 | | | |
| | *Borrelia* sp. Tick457 | | 4 | 847 | 7 | *Borrelia* sp. BR | 100 | EF141022.1 |
| | | | | | | *Borrelia* sp. strain | | |
| | | | | | | Mo063b-flaB | 100 | KY070335.1 |
| | | | | | | *Borrelia theileri* | 99 | KF569936.1 |
| | | | | MK049 | 24 | *Candidatus* Anaplasma | | |
| *Anaplasma* spp. | *Anaplasma* sp. Tick314 | 2 | 2 | 845 | 5 | boleense | 100 | KX987335.1 |
| *Anaplasma* | | | | MK049 | 24 | | | |
| *marginale* | *Anaplasma* sp. Tick283 | 2 | 2 | 844 | 4 | *Anaplasma marginale* | 100 | MH155593.1 |
| | | | | | | *Anaplasma centrale* | 100 | MF289482.1 |
| | | | | | | *Anaplasma ovis* | 100 | MG770440.1 |
| | | | | | | *Anaplasma capra* | 100 | MF000917.1 |
| | | | | | | *Anaplasma* | | |
| | | | | | | *phagocytophilum* | 100 | DQ648489.1 |
| | | | | MK049 | 24 | | | |
| *Ehrlichia* spp. | *Ehrlichia* sp. Tick428 | 2 | 2 | 849 | 6 | *Ehrlichia* spp. | 100 | KY594915.1* |
| | | | | | | *Ehrlichia canis* | 99 | KY594915.1 |
| | | | | | | *Ehrlichia ewingii* | 99 | U96436.1 |
| | | | | | | *Ehrlichia chaffeensis* | 99 | NR_074500.2 |
| | | | | | | *Ehrlichia muris* | 99 | KU535865.1 |
| | | | | | | *Ehrlichia minasensis* | 99 | NR_148800.1 |
| *Ehrlichia* | *Ehrlichia ruminantium* | | | MK049 | 20 | | | |
| *ruminantium* | Tick116 | 1 | 1 | 848 | 7 | *Erlichia ruminantium* | 100 | NR_074155.1 |
| *Babesia* | *Babesia bigemina* | | | MK071 | | | | |
| *bigemina* | Tick222 | 2 | 1 | 738 | 99 | *Babesia bigemina* | 100 | KP710227.1 |
| | | | | MK071 | 10 | | | |
| *Babesia bovis* | *Babesia bovis* Tick497 | 2 | 2 | 739 | 0 | *Babesia bovis* | 99 | AB367921.1 |

2.2.1. Detection of known TBPs in Caribbean ticks

Seven TBPs known to circulate in the Caribbean were detected in ticks from Guadeloupe and Martinique: *R. africae*, *E. ruminantium*, *An. marginale*, *B. bigemina*, *B. ovis*, *T. mutans* and *T. velifera* (Figure 2).

*Rickettsia* spp. were only detected in ticks collected in Guadeloupe (Figure 2). *R. africae* was identified in 95.6% of the *A. variegatum* samples (Figure 2). In contrast, *Rickettsia* spp. detected in 15.7-23.5% of the *R. microplus* samples from Guadeloupe were not directly identified as *R. africae* with the BioMarkTM system (Figure 2). Thus, 14 *A. variegatum* (6/14) and *R. microplus* (8/14) samples positive for *Rickettsia* spp. were tested by nested PCR with primers targeting the ompB gene; this was followed by sequencing. All the sequences recovered were identical and displayed 100% identity with *R. africae*, confirming that the *Rickettsia* spp. detected in *R. microplus* from Guadeloupe corresponded also to *R. africae*. (Table 2). The consensus sequence was deposited under the name *Rickettsia africae* Tick208 (accession number MK049851).

*E. ruminantium* was identified in 5.1% of the *A. variegatum* ticks from Guadeloupe (Figure 2). We confirmed the presence of *E. ruminantium* nucleic acids by testing one sample of *A. variegatum* by conventional PCR targeting the 16S rRNA genes; this was followed by amplicon sequencing. The sequence obtained displayed 100% sequence identity with *E. ruminantium* and was deposited under the name *Ehrlichia ruminantium* Tick116 (accession number MK049848) (Table 2).

*An. marginale* was identified in *R. microplus* ticks from both islands, with infection rates reaching 3.6-4.8% and 39.5-41.3% of specimens from Guadeloupe and Martinique, respectively (Figure 2). We confirmed the detection of *An. marginale* by testing two samples of *R. microplus* by conventional PCR targeting the 16S rRNA genes; this was followed by amplicon sequencing. We obtained two identical sequences, deposited under the name *Anaplasma* sp. Tick283 (accession number MK049844), which displayed 100% sequence identity with *Anaplasma* spp. including *An. marginale* (Table 2).

*B. bigemina* was detected in 0.6-1.2% and 12.5-12.8% of the *R. microplus* ticks from Guadeloupe and Martinique, respectively (Figure 2). *B. bovis* was only detected in ticks from Martinique, with an infection rate of 0.7% in *R. microplus* samples (Figure 2). As conventional and nested PCR did not succeed in detecting these parasites, we directly sequenced amplicons obtained with the *B. bigemina* and *B. bovis* designs developed here, and corresponding sequences were identified (accession numbers MK071738 and MK071739 respectively) (Table 2).

*T. velifera* and *T. mutans* were detected in both tick species and on both islands. *T. velifera* was identified in 42.3% of the *A. variegatum* samples and in 24.1-31.9% and 25.6-26% of the *R. microplus* samples from Guadeloupe and Martinique, respectively (Figure 2). Moreover, *T. mutans* was detected in 1.5% of the *A. variegatum* samples and in 1.8-2.4% and 1.4% of the *R. microplus* samples from Guadeloupe and Martinique, respectively (Figure 2). Unfortunately, neither conventional PCR nor BioMark amplicon sequencing succeeded in confirming the BioMark results.

#### 2.2.2. Detection of unexpected microorganisms in Caribbean ticks

Unexpected signals were obtained during the screening of microorganisms in ticks from Guadeloupe and Martinique, including the first detection of untargeted species belonging to the genera *Anaplasma*, *Ehrlichia*, *Borrelia* and *Leishmania* (Figure 2).

*Ehrlichia* spp. were detected in *R. microplus* ticks from both islands, with infection rates reaching 4.2-6.6% and 47.7-49.1% in Guadeloupe and Martinique, respectively (Figure 2). We tested two of the *Ehrlichia* spp.-positive *R. microplus* samples by conventional PCR targeting the 16S rRNA genes in order to identify the *Ehrlichia* spp. present in the Caribbean sample. We obtained two identical sequences, deposited under the name *Ehrlichia* sp. Tick428 (accession number MK049849) (Table 2). Phylogenetic and genetic distance analyses were performed using a portion of the 16S rRNA genes of several *Ehrlichia* species (Figure 3). The *Ehrlichia* sp. Tick428 sequence was found within a cluster including various uncharacterized *Ehrlichia* species detected in ticks from Asia and Africa (Figure 3).

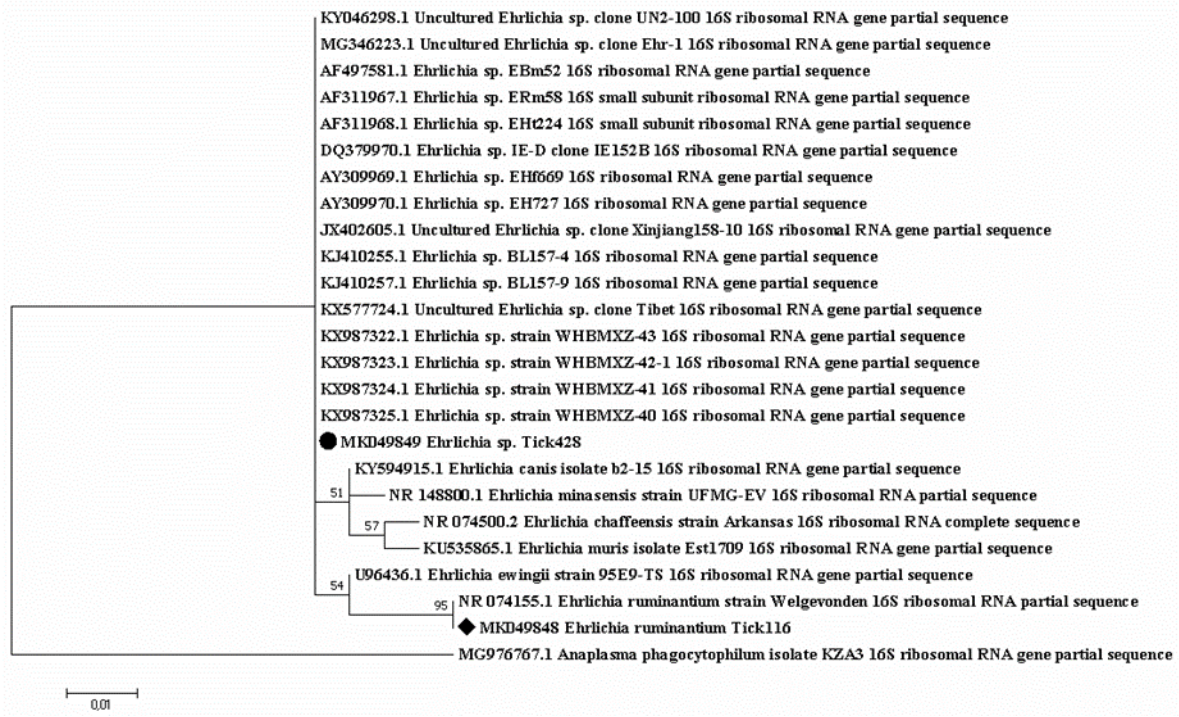



**Figure 3.** Phylogenetic analysis of 16S rRNA sequences of *Ehrlichia* spp. Phylogenetic analysis of 16S rRNA sequences of *Ehrlichia* spp. using the maximum likelihood method based on the Tamura-Nei model. In the phylogenetic tree, GenBank sequences, species designations and strain names are given. The sequences investigated in the present study are marked with a black circle (*Ehrlichia* sp. Tick428, accession number MK049849) and a black diamond (*Ehrlichia ruminantium* Tick116, accession number MK049848). The tree with the highest log likelihood (-413.76) is shown. The percentage of trees in which the associated taxa clustered together is shown above the branches (bootstrap values). The analysis involved 25 nucleotide sequences. There were a total of 206 positions in the final dataset.

In addition, in around 50% and 18% of the *R. microplus* specimens positive for *Anaplasma* spp., none of the *Anaplasma* species targeted by the BioMarkTM system gave signals, suggesting the presence of an unexpected or new *Anaplasma* spp. (Figure 2). We tested two of the *Anaplasma* spp.-positive *R. microplus* samples by conventional PCR targeting the 16S rRNA genes. We obtained two identical sequences, deposited under the name *Anaplasma* sp. Tick314 (accession number MK049845) (Table 2). This sequence displayed 100% sequence identity with *Candidatus* Anaplasma boleense. Phylogenetic and genetic distance analyses were performed using a portion of the 16S rRNA genes of several *Anaplasma* species (Figure 4). The *Anaplasma* sp. Tick314 sequence was found in a cluster including *Candidatus* Anaplasma boleense, *Anaplasma platys* and *Anaplasma phagocytophilum*.

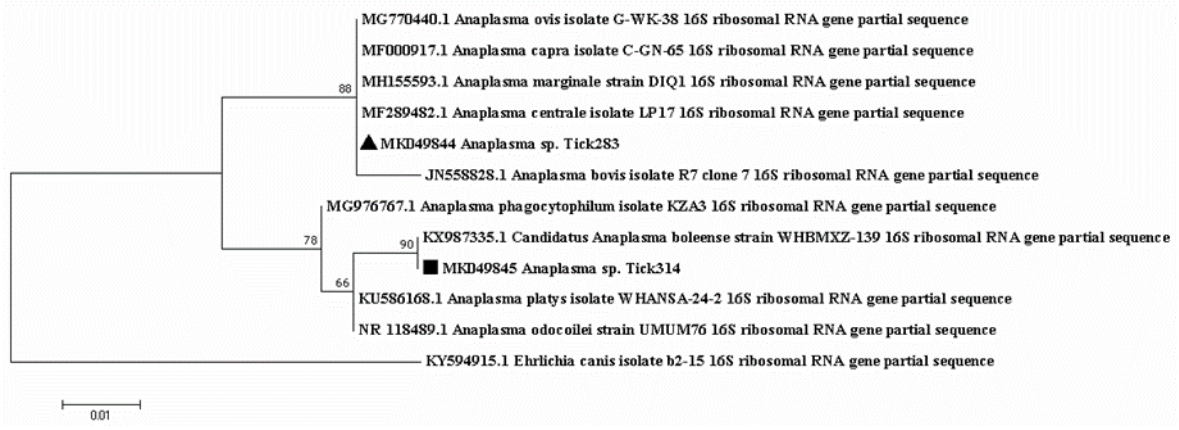



**Figure 4.** Phylogenetic analysis of 16S rRNA sequences of *Anaplasma* spp. Phylogenetic analysis of 16S rRNA sequences of *Anaplasma* spp. using the maximum likelihood method based on the Tamura-

Nei model. In the phylogenetic tree, GenBank sequences, species designations and strain names are given. The sequences investigated in the present study are marked with a black triangle (*Anaplasma* sp. Tick283, accession number MK049844) and a black square (*Anaplasma* sp. Tick314, accession number MK049845). The tree with the highest log likelihood (-473.44) is shown. The percentage of trees in which the associated taxa clustered together is shown above the branches (bootstrap values). The analysis involved 12 nucleotide sequences. There were a total of 243 positions in the final dataset.

*Borrelia* spp. were detected in both tick species from both islands (Figure 2). Infection rates reached 5.1% in *A. variegatum* and 0.6% and 4.3% in *R. microplus* from Guadeloupe and Martinique, respectively (Figure 2). None of the specific targeted *Borrelia* species causing Lyme disease (*Borrelia burgdorferi* sensu lato), or the *Borrelia* relapsing fever group, gave any positive results, suggesting the occurrence of a new or unexpected *Borrelia* spp. in our samples (Figure 2). We tested 30 of the *Borrelia* spp.-positive ticks by nested PCR targeting the flaB genes. Interestingly, we obtained two sequences according to the tick species analyzed. The *Borrelia* sp. Tick7 (accession number MK049846) sequence was recovered from one *A. variegatum* sample from Guadeloupe, and the *Borrelia* sp. Tick457 sequence (accession number MK049847) was recovered from four *R. microplus* samples from Martinique (Table 2). Phylogenetic and genetic distance analyses were performed using a portion of the flaB gene of several *Borrelia* species (Figure 5). Surprisingly, the *Borrelia* sp. Tick7 sequence recovered from the *A. variegatum* sample, and found to be closely related to *Bo. anserina*, displayed an intermediate position, sharing homology with both the relapsing fever and Lyme disease groups (Figure 5). Lastly, the *Borrelia* sp. Tick457 sequence recovered from the *R. microplus* samples confirmed the previous observations, forming a cluster with various relapsing fever *Borrelia* species encountered in hard ticks, including *Bo. lonestari* and *Bo. theileri* (Figure 5).

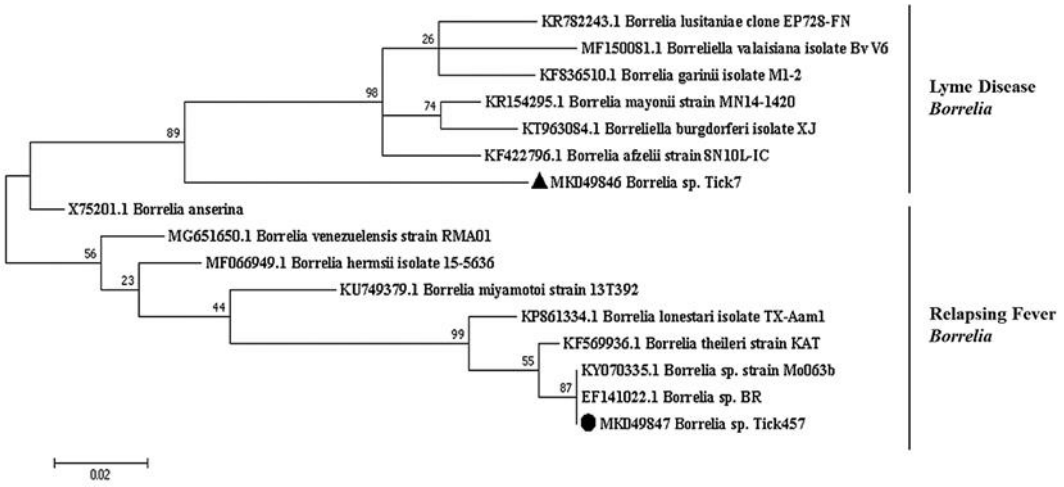



**Figure. 5.** Phylogenetic analysis of flaB sequences of *Borrelia* spp. Phylogenetic analysis of flaB sequences of *Borrelia* spp. using the maximum likelihood method based on the Tamura-Nei model. In the phylogenetic tree, GenBank sequences, species designations and strain names are given. The sequences investigated in the present study are marked with a black circle (*Borrelia* sp. Tick457, accession number MK049847) and a black triangle (*Borrelia* sp. Tick7, accession number MK049846). The Lyme disease and relapsing fever clades of *Borrelia* are marked. The tree with the highest log likelihood (-963.24) is shown. The percentage of trees in which the associated taxa clustered together is shown above the branches (bootstrap values). The analysis involved 16 nucleotide sequences. There were a total of 245 positions in the final dataset.

Lastly, 0.7% of the *R. microplus* ticks from Martinique were positive for *Leishmania* spp. (Figure 2). We tested two of the *Leishmania* spp.-positive ticks by nested PCR targeting the small sub-unit rRNA gene. We obtained one sequence from one sample, deposited under the name *Leishmania*

*martiniquensis* Tick389 (accession number MK049850) (Table 2). This sequence displayed 100% identity with both the *Leishmania martiniquensis* and *Leishmania siamensis* sequences (Table 2).

#### 2.2.3. Co-infections in ticks in Guadeloupe and Martinique

We analyzed the co-infections observed in *Amblyomma variegatum* (n=132 samples), *Rhipicephalus microplus* collected in Guadeloupe (n=116 samples, including individual and pooled specimens) and Martinique (n= 275 samples, including individual and pooled specimens). In Guadeloupe, almost all of the *A. variegatum* samples (99.2%) were infected with at least one pathogen whereas only 56% of the *R. microplus* samples were infected (Figure 6). In contrast, 81% of the *R. microplus* from Martinique were infected with at least one pathogen (Figure 6). High and similar percentages of the two tick species were infected with either one or two pathogens. The percentages drastically dropped for co-infection with three pathogens, with less than 10% of the ticks infected. Respectively one and nine *A. variegatum* and *R. microplus*, from Guadeloupe and Martinique, were co-infected with four pathogens and one *R. microplus* from Martinique was found infected with five pathogens (Figure 6).

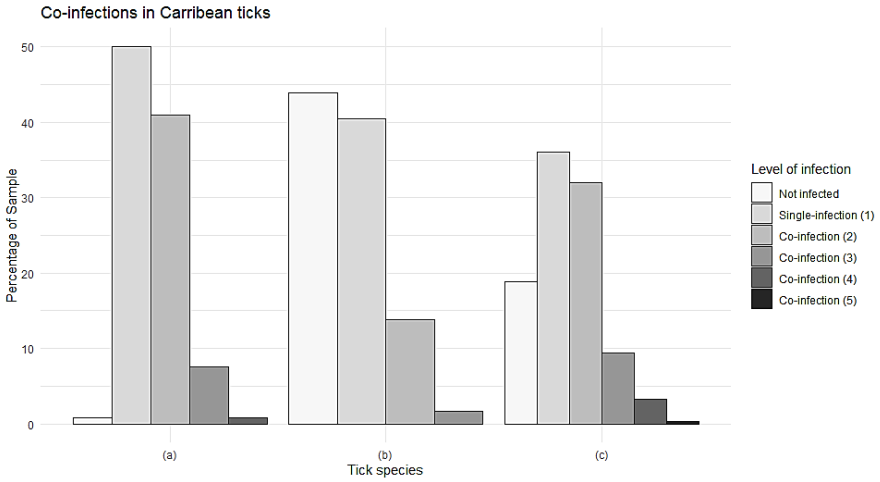



**Figure 6.** Co-infections detected in *Amblyomma variegatum* (n=132 samples) and *Rhipicephalus microplus* collected in Guadeloupe (n=116 samples) and Martinique (n=275 samples).

*A. variegatum* from Guadeloupe were find heavily infected by *R.africae*, yet it did not seem to affect the presence of other pathogen/microorganisms that were all find in co-infection with the bacteria (Table A3). Interestingly, in *R. microplus* from Guadeloupe, most of the single-infection reported corresponded to *R. africae* (12.9%) or *T. velifera* (21.6%) (Table A3). Positive association have been identified between *T. velifera* and *T. mutans*, and *Anaplasma* spp. / *Borrelia* spp. (Table A5). Finally, in *R. microplus* from Martinique, five positive associations have been detected, including *T. mutans* / *T. velifera*, *T. mutans* / *Leishmania* spp., *T. mutans* / *Borrelia* spp., *T. velifera* / *B. bigemina*, *A. marginale*/ *Ehrlichia* spp. (Table A6). The result of the co-occurrence test should be taken with cautions and deserve further investigation regarding the few number of positive samples (Table A5-A6). Nevertheless, no exclusion seemed to occur between the pathogens/microorganisms detected in the two tick species from Guadeloupe and Martinique. More details on co-infections in ticks from Guadeloupe and Martinique are available in Appendix B.

## 3. Discussion

In this study, a high-throughput microfluidic real-time PCR system based on the use of multiple primers/probes was developed for large-scale surveys of bacteria and protozoans potentially transmitted by ticks from the Caribbean area. The association of genus and species primer/probe designs targeting TBPs improved the technology's screening capacity, enabling not only the identification of infectious agents known to circulate in the studied area, but also the detection of

unsuspected TBPs and new microorganisms belonging to the main bacterial and protozoan genera/phyla involved in TBDs worldwide. Nevertheless, as some endosymbiotic microorganisms may belong to known TBP genera, such as *Rickettsia* and *Coxiella*, confirmatory tests are required before suggesting the presence of a pathogenic microorganism [23-25]. When analyzing the specificity of the microfluidic real-time PCR system, cross-reactions were observed for some designs targeting closely related species; these must be taken into account when interpreting the results. Due to high design constraints and a lack of available sequences in public databases, the improvement of such cross-reacting oligonucleotides remains challenging. Here, the concomitant use of bacterial and protozoan genera can assist in identifying non-specific signals. In addition to detecting microorganisms, we developed sets of primers and probes enabling the molecular identification of the three main tick species involved in TBDs in the Caribbean: *A. variegatum*, *R. microplus* and *R. sanguineus* s.l. As the morphological identification of ticks collected in the field remains challenging, molecular identification can be used to confirm the identification of the tick species analyzed.

We used the newly developed high-throughput microfluidic real-time PCR system to perform an exploratory epidemiological study of TBPs and microorganisms potentially circulating in Caribbean ticks. The analysis provided an overview of the diversity of microorganisms belonging to the main bacterial and protozoan genera potentially transmitted by ticks. It enabled the detection both of known TBPs of public and animal health importance in the area that require surveillance and of unexpected microorganisms occurring in Caribbean ticks.

The four main pathogens responsible for ruminant diseases in the Caribbean – currently classified as notifiable diseases by the World Organisation for Animal Health (OIE) – have been detected by the microfluidic real-time PCR system. These are *E. ruminantium* in *A. variegatum* specimens and *An. marginale*, *B. bigemina* and *B. bovis* in *R. microplus*.

Interestingly, the *E. ruminantium* infection rate in *A. variegatum* reported in our study was much lower compared to in previous studies conducted between 2003 and 2005 in Guadeloupe (5.1% versus 36.7%) [12]. Although different study designs were used (different sampling strategies, study periods, detection methods, etc.), which may explain this difference, it would be worth further investigating whether the tick infection rate for *E. ruminantium* has decreased in Guadeloupe and possibly assessing the epidemiological impact in terms of the incidence and prevalence of heartwater in the ruminant population. These results are all the more surprising since systematic TBT surveillance and control programs have been discontinued following the end of the POSEIDOM eradication programs in 2006.

In this study, we have documented infection rates for *B. bigemina*, *B. bovis* and *An. marginale* in the *R. microplus* vector tick in the French West Indies for the first time. Indeed, records of such pathogens are mostly based on seroprevalence studies in cattle [7,8,10].

*R. microplus* ticks are both vectors and reservoirs of *B. bigemina* and *B. bovis*, transmitting the parasites transovarially and trans-stadially [26,27]. As *R. microplus* ticks and cattle are both reservoirs of infection, the infection rates reported here seemed quite low. The life cycle of *Babesia* spp. requires complex interactions with its two hosts, which are the tick vector and the vertebrate host. The efficiency of tick acquisition and of transovarial and trans-stadial transmission of *B. bovis* and *B. bigemina* by *R. microplus*, involved in the long-term persistence of *Babesia* spp. in nature, is still poorly understood and warrants further investigations [26,27].

Interestingly, *An. marginale* was detected in *R. microplus* from both islands, but the infection rate reported in ticks from Guadeloupe seemed lower compared to in Martinique. The same trend had been reported during previous seroprevalence studies [7,8,10]. Anaplasmosis can be transmitted by vectors other than ticks, and some cattle breeds are known to be more susceptible than others to *Anaplasma* infection [10]. The difference in *Anaplasma* infection rate in ticks between the two islands may have been due to differences in the cattle populations. Indeed, there are mainly local Creole and mixed European-Creole breeds in Guadeloupe. These are known to be more resistant to anaplasmosis than Brahman and European breeds, which are the main breeds reared in Martinique [10]. In addition, other factors, including differences in the population dynamics of alternate vectors such as flies, may also have contributed to this difference.

Among the other known TBPs detected, we also found pathogens with low health impact in the Caribbean – almost considered as endosymbionts – such as *R. africae*, *T. velifera* and *T. mutans* in their *A. variegatum* vector and surprisingly in *R. microplus* ticks.

With almost all of the *A. variegatum* found to be infected, the *R. africae* infection rate was the highest ever reported in the Caribbean [9,13,14,28]. As *A. variegatum* is both the vector and the reservoir of the pathogen, with transovarial and trans-stadial transmission rates reaching 100%, this high level of *R. africae* infection is not surprising per se [14,29]. Interestingly, the high *R. africae* infection rate in vector ticks, associated with a very low number of African tick-bite fever cases in the Caribbean, highlights the difficulty, in some cases, of clearly distinguishing between endosymbiosis and pathogenicity [9,15]. The biological relationship between *R. africae* and *A. variegatum* as well as the strain variety and virulence of *R. africae* in the Caribbean should be investigated in order to better assess risks and guide prevention measures, especially for travelers [23,24,30]. The absence of direct identification of *R. africae* in *R. microplus* ticks was probably due to lower sensitivity of the specific target design compared to the genus target design. Indeed, *Rickettsia* spp.-positive *R. microplus* samples displayed rather high Ct values, suggesting a low infection level that may have been below the detection limit for *R. africae*. The unusual presence of *R. africae* in *R. microplus* ticks may have been due to the co-occurrence of the two tick species, *R. microplus* and *A. variegatum*, on cattle. As the ticks here were collected partially engorged, the presence of *R. africae* in *R. microplus* may have been due to bacteria circulating in cattle blood picked up by engorging ticks, or to cross-contamination with *R. microplus* ticks co-feeding next to infected *A. variegatum* [31,32].

This study provides the first update on the detection of *T. mutans* and *T. velifera* in Caribbean ticks. Indeed, references to these parasites in the Caribbean are relatively old, and no prevalence studies have been conducted since, whether in ticks or in cattle [5,6,33]. The low pathogenicity of these piroplasms may explain the lack of diagnoses and the scarcity of information available on their distribution and prevalence in the Caribbean. However, these parasite species may play an important role in theileriosis management and protection, as chronically infected cattle can develop immunity and heterologous protection against other pathogenic *Theileria* species, such as *Theileria parva* [34]. Unfortunately, we did not succeed in confirming these results by conventional or nested PCR, suggesting either a level of infection below the detection threshold, or simply false signals.

Lastly, the high-throughput microfluidic real-time PCR system enabled the detection of unexpected and/or potentially new microorganisms, leading to the recovery of nucleotide sequences of *Anaplasma* spp., *Ehrlichia* spp., *Borrelia* spp. and *Leishmania* spp. in ticks collected in Guadeloupe and Martinique.

The *Ehrlichia* sp. Tick428 sequence detected here formed a cluster with other uncharacterized Ehrlichia species detected in ticks from Asia and Africa [13,35-39]. However, given the highly conserved nature of the 16S rRNA genes, we could not more accurately define phylogenetic relationships within the *Ehrlichia* species group. The *Anaplasma* sp. Tick314 sequence was identified as *Candidatus* Anaplasma boleense, a bacterium described in ticks and mosquitoes in China [38,40]. No further information is available regarding the epidemiology of *Candidatus* Anaplasma boleense. These observations highlight the need to set up characterization studies. Indeed, high-throughput detection technologies can highlight the presence of DNA from potentially new microorganisms, but it will still be necessary to isolate and characterize them in order to first confirm their existence and then determine whether their presence in ticks poses a risk to public or animal health.

Here we provided the first report of *Borrelia* spp. in ticks from Guadeloupe and Martinique. Two different sequences were recovered, according to the tick species analyzed. In *A. variegatum*, a sequence named *Borrelia* sp. Tick7 was detected and was closely related to *Bo. anserina*, the agent of avian spirochetosis. Both of them seemed to define an intermediate position between the relapsing fever and Lyme disease groups. In contrast, the *Borrelia* sp. Tick457 sequence found in *R. microplus* sample, clustered with uncharacterized *Borrelia* spp. described *R. microplus* specimens from Madagascar and Brazil, such as *Borrelia* sp. strain Mo063b and *Borrelia* sp. BR, and with relapsing fever Borrelia species encountered in hard ticks, including *Borrelia lonestari* and *Bo. theileri* [41,42]. Interestingly, the same observations had recently been made regarding *Borrelia* spp. found in *A.*

*variegatum* and *R. microplus* ticks from Ethiopia and Côte d'Ivoire [43,44]. As *A. variegatum* and *R. microplus* were imported into the Caribbean from Africa during the time of the Atlantic triangular trade, we may have detected bacteria probably characterized by an old introduction through infected ticks and subsequent local evolution within their vector over a long period [4,45]. *Borrelia* spp. and borreliosis case reports in the Caribbean are scarce and still being debated. In Cuba, one study suggested the presence of antibodies to *Borrelia burgdorferi* sensu stricto in human sera associated with clinical cases of Lyme disease-like syndrome [46,47]. However, the real specificity of these serum antibodies has been questioned [48]. In the US Virgin Islands, seropositivity for *Borrelia hermsii* and closely related species was reported in association with a human case of relapsing fever [49]. Lastly, erythema migrans-like skin lesions and illness were reported in four Caribbean nationals [50]. Regarding the importance of *Borrelia* spp. for human and animal health, the characterization of these potential new *Borrelia* species that seemed associated with tropical tick species requires further investigation.

Lastly, *Leishmania* spp. were detected in *R. microplus* specimens from Martinique, and one sequence was identified as *Leishmania martiniquensis* Tick389 (accession number MK049850). Studies on *Leishmania* nomenclature have highlighted the fact that isolates of "*L. siamensis*" have never been officially characterized and that therefore, this name should not be used [51-54]. Thus, since all the sequences – except one – reported as "*L. siamensis*" in databases should be considered as synonyms of *L. martiniquensis*, we assumed the occurrence of *L. martiniquensis* here. Parasites of the genus *Leishmania* are usually transmitted by female phlebotomine sand flies (Diptera: Psychodidae: Phlebotominae) and generally involve a wide variety of animal species, mainly including dogs and canids in the epidemiological cycle. They are responsible for leishmaniasis, a zoonosis widespread in tropical and sub-tropical areas [54]. *L. martiniquensis* belongs to the *L. enriettii* complex and has been described in Martinique and Thailand, where it was responsible for both cutaneous and visceral leishmaniosis [51,54-56]. *L. martiniquensis* is suspected to be endemic in Martinique [55]. Although phlebotomines and rodents are present in Martinique, neither vectors nor reservoirs of this parasite have yet been described [55]. Our study represents the first report of *L. martiniquensis* in *R. microplus* ticks from the French West Indies. Although *Leishmania* spp. have been reported in ticks (*L. infantum* in *R. sanguineus s.l.*, and *L. guyanensis* in *R. microplus* ticks in Peru, for example), the role of ticks in *Leishmania* transmission is still being debated, and no evidence of vector capacity has been reported yet [57-59]. Moreover, the finding of *Leishmania* spp. in a tick species that feeds mainly on cattle also raises questions about the potential role of cattle in the epidemiology of leishmaniasis [60,61]. The participation of ticks in *Leishmania* epidemiology warrants further investigation, especially since *R. microplus* ticks could parasitize humans [62].

Surprisingly, co-infections with two or more TBPs were found in more than 50% of the infected ticks, both for *A. variegatum* and *R. microplus* and on the two islands. In addition, we could not identify any exclusion of infection between pathogens. These observations illustrate the efficiency of ticks as reservoirs of multiple pathogens with no apparent significant effects on their life traits.

To conclude, although screening tools are useful for the discovery of pathogens in ticks, the epidemiological significance of such results warrants further analysis. Detecting a microorganism's DNA in ticks, especially in partially engorged ticks, does not necessarily mean that the ticks are involved in the microorganism's life cycle; however, it provides useful information to supplement vector competence studies [16]. Nevertheless, the detection of potentially new microorganisms in ticks from the French West Indies has opened up new research perspectives for the future on the epidemiology of TBPs in the Caribbean. A region-wide epidemiological survey on TBPs in ticks collected in different countries and territories of the Caribbean area, organized in collaboration with the Caribbean Animal Health Network (CaribVET) in order to strengthen our results, may be an interesting way to supplement and strengthen some of this paper's findings.

## 4. Materials and Methods

### *4.1 Ticks collected in Guadeloupe and Martinique*

The ticks used in this study were collected as part of two separate epidemiological surveys conducted in Guadeloupe (between February 2014 and January 2015) and Martinique (between February and March 2015), respectively. In Guadeloupe, adult ticks (any species, any sex) were collected from 40 cattle originating from 22 different herds that were sampled in nine localities situated in six different biotopes (urban area, dry coastal regions, valleys and hills, evergreen seasonal forest, sub-mountainous rainforest, swamp forest). In Martinique, engorged females of *R. microplus* only were collected from cattle in 29 farms participating in a study on acaricide resistance in ticks. All the ticks were collected from cattle with the permission of farmers and cattle owners. The ticks were morphologically identified at species level [63]. A total of 578 adult ticks were included in the study: 132 *A. variegatum* and 165 *R. microplus* ticks from Guadeloupe and 281 *R. microplus* ticks from Martinique (see maps, Figure 2). The GPS coordinates of the tick collection sites are available in Table S1. All the ticks were partially engorged, and then stored at -80°C.

### *4.2 DNA extraction of ticks collected in Guadeloupe and Martinique*

For 20 mg of tick, 1 ml of recently prepared PBS 1X was added to the sample. The ticks were then washed by gently shaking for 2-3 min at 7 Hz/s in a TissueLyser (Qiagen, Germany). After discarding the supernatant, the ticks were frozen at -80°C for 15-20 min. A steel ball was then added and the samples were crushed twice for 2 min at 30 Hz/s with the TissueLyser (Qiagen, Germany). 450 µl of fresh PBS 1X were added to the samples. The samples were vortexed for 10 s and then centrifuged for 2-3 min at 8000 g. Lastly, 20 µl of Proteinase K were added to 180 µl of crushed tick sample and DNA was extracted using the NucleoSpin® 96 Virus Core Kit (Macherey-Nagel, Germany) and the Biomek4000 automated platform (Beckman Coulter). This protocol enables the simultaneous extraction of both DNA and RNA. Total nucleic acid per sample was eluted in 160 µl of rehydration solution and stored at -80°C until further use. *A. variegatum* ticks were individually extracted. *R. microplus* ticks were extracted both individually and in pools of two to four adult specimens when they were too small to be treated individually.

### *4.3 Assay design*

The list of pathogens to be monitored, the sets of primers and probes required for their detection, as well as the targeted genes are shown in Table 1. Some of the oligonucleotides were specifically designed for the purposes of this study; the others came from Michelet et al., 2014 [18]. The newly developed oligonucleotides were validated for a range of dilutions of positive controls, including cultures, plasmids and DNA samples (Table 1, Table S2), by real-time TaqMan PCR assays on a LightCycler® 480 (LC480) (Roche Applied Science, Germany). Real-time PCR assays were performed with LightCycler® 480 Probe Master Mix 1× (Roche Applied Science, Germany), using 200 nM of primers and probes in a final volume of 12 µl, and 2 µl of control DNA were added. The thermal cycling program was as follows: 95°C for 5 min, 45 cycles at 95°C for 10 s and 60°C for 15 s, and one final cooling cycle at 40°C for 10 s.

### *4.4 Pre-amplification of DNA samples*

All the DNA samples were subject to pre-amplification in order to enrich the pathogenic DNA content compared with tick DNA. PerfeCTa® PreAmp SuperMix (Quanta Biosciences, Beverly, USA) was used for DNA pre-amplification, following the manufacturer's instructions. All the primers were pooled (except those targeting the tick species), with a final and equal concentration of 45 nM each. The pre-amplification reaction was performed in a final volume of 5 µl containing 1 µl of PerfeCTa PreAmp SuperMix (5X), 1.25 µl of pooled primer mix, 1.25 µl of DNA and 1.5 µl of Milli-Q water, with one cycle at 95°C for 2 min and 14 cycles at 95°C for 10 s and 60°C for 3 min. At the end of the cycling program, the reactions were 1:10 diluted. The pre-amplified DNA were stored at -20°C until use.

### *4.5 High-throughput microfluidic real-time PCR*

High-throughput microfluidic real-time PCR amplifications were performed using the BioMark™ real-time PCR system (Fluidigm, USA) and 96.96 dynamic arrays (Fluidigm, USA), enabling up to 9,216 individual reactions to be performed in one run [18]. Real-time PCRs were performed using 6-carboxyfluorescein (6-FAM)- and Black Hole Quencher (BHQ1)-labeled TaqMan probes with TaqMan Gene Expression Master Mix (Applied Biosystems, USA) following the manufacturer's instructions. The cycling conditions were as follows: 2 min at 50°C and 10 min at 95°C, followed by 40 cycles of two-step amplification for 15 s at 95°C and 1 min at 60°C. The BioMark™ real-time PCR system was used for data acquisition and the Fluidigm real-time PCR analysis software for Ct value determination. Three kinds of controls per chip were used for experiment validation: a negative water control to exclude contamination; a DNA extraction control, corresponding to primers and probes targeting a portion of the 16S rRNA gene of ticks; and an internal control, to check the presence of PCR inhibitors made of DNA from Escherichia coli strain EDL933, added to each sample with specific primers and probes targeting the eae gene [64]. For the relative specificity analysis of the newly designed Biomark system, DNA of 62 positive controls were used as template (Table S2). Then, for the epidemiological survey of TBPs in Caribbean ticks, the 523 DNA samples of *A. variegatum* and *R. microplus* from Guadeloupe and Martinique were used as template.

### *4.6 Infection rates for ticks from the French West Indies*

Depending on the tick species and the island of origin, for each detected pathogen, infection rates (the proportion of infected ticks divided by the total number of ticks analyzed) were calculated. The majority of the samples were single specimens of ticks. When ticks were too small to be treated individually, they were grouped into pools of two to four specimens. Thus, of the 523 samples analyzed, 47 consisted of a pool of two to four tick specimens. The final estimation of infection rates also includes the pools and is therefore expressed as the minimum (assuming at least one positive tick in the pools) and maximum (assuming all positive ticks in the pools) proportions of infected ticks out of the total number of ticks analyzed.

### *4.7 PCRs and sequencing for the confirmation of results*

Conventional PCRs/nested PCRs using primers targeting different genes or regions than those of the BioMark™ system were used to confirm the presence of pathogenic DNA in some field samples and positive controls (Table 3). Amplicons were sequenced by Eurofins MWG Operon (BIOMNIS-EUROFINS GENOMICS, France) and then assembled using BioEdit software (Ibis Biosciences, Carlsbad, CA, USA). An online BLAST (Basic Local Alignment Search Tool) search was used to compare the nucleotide sequences found in this study to reference sequences listed in GenBank sequence databases (NCBI).

**Table 3.** Primers used to confirm the presence of pathogenic DNA in tick samples, and positive controls.

| Pathogen | Targeted gene | Primer name | Sequence (5' → 3') | Length (bp) | References |
|---|---|---|---|---|---|
| Rickettsia spp. | gltA | Rsfg877 | GGGGGCCTGCTCACGGCGG | 381 | [65] |
| | | Rsfg1258 | ATTGCAAAAAGTACAGTGAACA | | |
| | ompB | Rc.rompB.4362p | GTCAGCGTTACTTCTTCGATGC | 475 | [66] |
| | | Rc.rompB.4,836n | CCGTACTCCATCTTAGCATCAG | | |
| | | Rc.rompB.4,496p | CCAATGGCAGGACTTAGCTACT | 267 | |
| | | Rc.rompB.4,762n | AGGCTGGCTGATACACGGAGTAA | | |
| Anaplasma/Ehrlichia spp. | 16S rRNA | EHR16SD | GGTACCYACAGAAGAAGTCC | 345 | [67] |
| | | EHR16SR | TAGCACTCATCGTTTACAGC | | |

| | | | | | |
|---|---|---|---|---|---|
| Borrelia spp. | flaB | FlaB280F | GCAGTTCARTCAGGTAACGG | 645 | [68] |
| | | FlaRL | GCAATCATAGCCATTGCAGATTGT | | |
| | | FlaB737F | GCATCAACTGTRGTTGTAACATTA ACAGG | 407 | |
| | | FlaLL | ACATATTCAGATGCAGACAGAGG T | | |
| Leishmania spp. | SSU rRNA | R221 | GGTTCCTTTCCTGATTTACG | 603 | [69] |
| | | R332 | GGCCGGTAAAGGCCGAATAG | | |
| | | R223 | TCCATCGCAACCTCGGTT | 358 | |
| | | R333 | AAAGCGGGCGCGGTGCTG | | |

### *4.8 Phylogenetic sequence analysis*

Alignments were performed using ClustalW [70]. Maximum likelihood trees were generated by 1,000 bootstrap repetitions based on the Tamura-Nei model [71] in MEGA7 [72]. The initial tree(s) for the heuristic search were obtained automatically by applying neighbor-joining and BioNJ algorithms to a matrix of pairwise distances estimated using the maximum composite likelihood (MCL) approach and then selecting the topology with superior log likelihood value. The tree was drawn to scale, with branch lengths measured in the number of substitutions per site. The codon positions included were 1st+2nd+3rd+Noncoding. All positions containing gaps and missing data were eliminated. Further information is provided in the figure legends.

## 5. Conclusions

Our study demonstrated the high ability of microfluidic real-time PCR technology to provide a rapid overview of the diversity of TBPs of veterinary and medical importance present in ticks from the Caribbean. This innovative high-throughput tool is promising and could significantly improve the surveillance and exploration of TBPs, enabling the rapid screening of multiple microorganisms especially in regions where few epidemiological data are available and TBDs are numerous.

**Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1, Table S1: GPS coordinates of the tick collection sites and number of ticks collected. A total of 578 adult ticks collected from cattle from Guadeloupe and Martinique were used for the screening of tick-borne pathogens with the newly implemented BioMark™ real-time PCR system, Table S2: List of the positive control samples used for the relative specificity analysis of the Biomark system developped in this study, Figure S1: Improvement of detection signals by pre-amplification. Test of primer/probe set sensitivity for a range of dilutions of positive controls by TaqMan real-time PCR using LightCycler 480, before and after pre-amplification. Results of the sensitivity test of the *Leishmania infantum* design using a *Leishmania infantum* culture, before (a) and after (c) pre-amplification; Results of the sensitivity test of the *Rickettsia* spp. design using *Rickettsia conorii*-positive controls (extracted from an infected *Rhipicephalus sanguineus* sensu lato tick), before (b) and after (d) pre-amplification.

**Author Contributions:** All authors have read and agree to the published version of the manuscript. Conceptualization, E.A., J.P. and S.M.; methodology, M.G., E.D., V.P., R.A., C.G., S.D. and M.V.T.; formal analysis, M.G.; investigation, M.G.; writing—original draft preparation, M.G.; writing—review and editing, M.V.T., E.A., J.P. and S.M.; supervision, E.A., J.P. and S.M.; funding acquisition, M.V.T., E.A., J.P. and S.M.

**Funding:** This research was funded by grants from the French Agency for Food, Environmental and Occupational Health & Safety (ANSES, CoVetLab grant), the French Agricultural Research Centre for International Development (CIRAD) and the French National Institute for Agricultural Research (INRA, PATHO-ID metaprogram). This study was partly supported by the MALIN project on "Surveillance, diagnosis, control and impact of infectious diseases of humans, animals and plants in tropical islands", supported by the European Union, in the framework of the European Regional Development Fund (ERDF), and the Regional Council of Guadeloupe. The study used ticks collected in Guadeloupe and Martinique as part of previous projects: the PathoID "Rodent and tick pathobiome" project funded by INRA and the ResisT project on "Assessment of tick resistance to acaricides in the Caribbean – Development of strategies to improve surveillance and control of tick-borne diseases in ruminants", funded by the Regional Cooperation Fund (FCR) of Guadeloupe and Martinique. This work was performed in the framework of EurNegVec COST Action TD1303.

**Acknowledgments:** In this section you can acknowledge any support given which is not covered by the author contribution or funding sections. This may include administrative and technical support, or donations in kind (e.g., materials used for experiments).

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

## Appendix A

Analysis of the relative specificity of the 61 sets of primers and probe constituting the Biomark system developed in this study.

In order to identify potential cross-reactions, we analyzed the relative specificity of the 61 sets of primers and probe constituting the BioMark system used in this study using 62 positive control samples including DNA from bacterial or parasitic cultures, or DNA from tick or blood samples known to be infected, or plasmidic constructions (see Table S2).

Of the 61 designs, 42 designs were specific of their target. The Tick spp. design, used as a tick nucleic acid extraction control, was able to detect *A. variegatum* and *R. sanguineus* s.l. samples as well as the DNA of the *R. sanguineus* s.l. tick present in the *Rickettsia conorii* positive control as expected (Table A1). However, the DNA of ticks from the *R. microplus* control sample and other positive controls including tick DNA (such as the *Borrelia lonestari*, *Anaplasma phagocytophilum* controls, etc.) were not detected (Table A1). The detection ability of this design was corrected by adding the Tick spp. primers during the pre-amplification step; these had initially been excluded since the objective was to enrich pathogenic DNA content compared to tick DNA (data not shown). Eight designs displayed cross-reactions with one to two closely related species, and seven designs displayed unexpected signals corresponding likely to the detection of unexpected co-infection in complex control samples such as DNA extracted from ticks or blood samples (Table A1). Finally three design were removed from the system, one design due to a lack of efficiency (no detection of the target), and two designs were not specific, displaying multiple cross-reactions (Table A1).

**Table A1.** List of designs and their specificity using the BioMark system. CR: cross-reactions with closely related species samples; CI: potential co-infections in control samples.

| Design | Target detection | Specificity | Outgroup control samples |
|---|---|---|---|
| *Rickettsia spp. gltA* | *YES* | CI | *Borrelia lonestari* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Ehrlichia chaffensis* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Ehrlichia ewingii* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Amblyomma variegatum* (Wild tick) |
| *Rickettsia massiliae 23S-5S ITS* | *YES* | *CR* | *Rickettsia slovaca* (Culture) |
| *Rickettsia rickettsii 23S-5S ITS* | *YES* | *CR* [1] | *Rickettsia slovaca* (Culture) |
| | *YES* | | *Rickettsia conorii* (Infected *R. sanguineus* s.l. ticks) |
| | *YES* | | *Rickettsia africae* (Culture) |
| | *YES* | | *Amblyomma variegatum* (Wild tick) |
| *Rickettsia conorii sca1* | *YES* | *CR* | *Rickettsia slovaca* (Culture) |
| *Rickettsia africae sca1* | *YES* | CI | *Amblyomma variegatum* (Wild tick) |
| *Rickettsia felis orfB* | *YES* | CI | *Borrelia lonestari* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Ehrlichia chaffensis* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Ehrlichia ewingii* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) |
| *Rickettsia typhi ompB* | *YES* | *YES* | |

| | | | |
|---|---|---|---|
| *Rickettsia prowazekii gltA* | *YES* | *YES* | |
| *Borrelia spp. 23S rRNA* | *YES* | *YES* | |
| *Borrelia burgdorferi* sensu stricto *glpA* | *NO* [1] | | |
| *Borrelia anserina fla* | *YES* | *YES* | |
| *Borrelia lonestari glpQ* | *YES* | *YES* | |
| *Borrelia parkeri gyrB* | *YES* | *YES* | |
| *Borrelia bissettii rpoB* | *YES* | *YES* | |
| *Borrelia theileri glpQ* | *YES* | *YES* | |
| *Bartonella spp. ssrA* | *YES* | *YES* | |
| *Bartonella bacilliformis rpoB* | *YES* | *CR* | *Bartonella henselae* (Culture) |
| *Bartonella henselae ribC* | *YES* | *CR* | *Bartonella bacilliformis* (Culture) |
| *Bartonella vinsonii subsp. berkhoffii ITS* | *YES* | *YES* | |
| *Coxiella burnetii icd* | *YES* | *YES* | |
| *Coxiella burnetii IS 1111* | *YES* | *YES* | |
| *Francisella tularensis tul4* | *YES* | *YES* | |
| *Francisella tularensis fopA* | *YES* | *YES* | |
| *Anaplasma spp. 16S rRNA* | *YES* | *YES* | |
| *Anaplasma marginale msp1b* | *YES* | *YES* | |
| *Anaplasma phagocytophilum msp2* | *YES* | *YES* | |
| *Anaplasma platys groEL* | *YES* | *YES* | |
| *Anaplasma bovis groEL* | *YES* | *YES* | |
| *Anaplasma ovis msp4* | *YES* | *YES* | |
| *Ehrlichia spp. 16S rRNA* | *YES* | *YES* | |
| *Ehrlichia canis gltA* | *YES* | *YES* | |
| *Ehrlichia chaffeensis dsb* | *YES* | *YES* | |
| *Ehrlichia ewingii dsb* | *YES* | *YES* | |
| *Ehrlichia ruminantium gltA* | *YES* | *YES* | |
| *Panola Mountain Ehrlichia gltA* | *YES* | *YES* | |
| *Neoehrlichia mikurensis groEL* | *YES* | *YES* | |
| *Aegyptianella pullorum groEL* | *YES* | *YES* | |
| *Apicomplexa 18S rRNA* | *YES* | CI | *Borrelia lonestari* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Anaplasma marginale* (Infected cow blood) |
| | *YES* | CI | *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) |
| | *YES* | CI | *Neoehrlichia mikurensis* (Infected rodent blood) |
| | *YES* | CI | *Rhipicephalus sanguineus* s.l. (Wild tick) |
| *Babesia canis vogeli hsp70* | *YES* | *CR* | *Babesia canis canis* (Infected dog blood) |
| *Babesia ovis 18S rRNA* | *YES* | *CR* [1] | *Rickettsia massiliae* (Culture) |
| | *YES* | | *Borrelia lonestari* (Infected *A. americanum* ticks) |
| | *YES* | | *Anaplasma marginale* (Infected cow blood) |
| | *YES* | | *Ehrlichia chaffensis* (Infected *A. americanum* ticks) |
| | *YES* | | *Ehrlichia ewingii* (Infected *A. americanum* ticks) |
| | *YES* | | *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) |
| | *YES* | | *Neoehrlichia mikurensis* (Infected rodent blood) |

| | | | |
|---|---|---|---|
| | *YES* | | *Babesia divergens* (Culture) |
| | *YES* | | *Babesia canis rossi* (Infected dog blood) |
| | *YES* | | *Babesia canis canis* (Infected dog blood) |
| | *YES* | | *Babesia canis vogeli* (Infected dog blood) |
| | *YES* | | *Babesia microti* (Culture) |
| | *YES* | | *Theileria annulata* (Culture) |
| | *YES* | | *Theileria lestoquardi* (Culture) |
| | *YES* | | *Theileria parva* (Culture) |
| | *YES* | | *Hepatozoon canis* (Infected dog blood) |
| | *YES* | | *Rhipicephalus sanguineus* s.l. (Wild tick) |
| *Babesia bigemina 18S rRNA* | *YES* | CI | *Anaplasma marginale* (Infected cow blood) |
| *Babesia gibsoni Rap1* | *YES* | *YES* | |
| *Babesia caballi Rap1* | *YES* | *YES* | |
| *Babesia bovis CCTeta* | *YES* | *YES* | |
| *Babesia duncani ITS2* | *YES* | *YES* | |
| *Babesia microti CCTeta* | *YES* | *CR* | *Babesia canis canis* (Infected dog blood) |
| *Theileria parva 18S rRNA* | *YES* | *CR* | *Theileria annulata* (Culture) |
| | *YES* | *CR* | *Theileria lestoquardi* (Culture) |
| *Theileria mutans ITS* | *YES* | *YES* | |
| *Theileria velifera 18S rRNA* | *YES* | *YES* | |
| *Theileria equi ema1* | *YES* | *YES* | |
| *Cytauxzoon felis ITS2* | *YES* | *YES* | |
| *Hepatozoon spp 18S rRNA* | *YES* | CI | *Neoehrlichia mikurensis* (Infected rodent blood) |
| | *YES* | CI | *Rhipicephalus sanguineus* s.l. (Wild tick) |
| *Hepatozoon canis 18S rRNA* | *YES* | CI | *Rhipicephalus sanguineus* s.l. (Wild tick) |
| *Hepatozoon americanum 18S rRNA* | *YES* | CR | *Hepatozoon canis* (Infected dog blood) |
| | *YES* | CI | *Rhipicephalus sanguineus* s.l. (Wild tick) |
| *Leishmania spp hsp70* | *YES* | *YES* | |
| *Leishmania infantum ITS* | *YES* | *YES* | |
| *Rangelia vitalii 18S rRNA* | *YES* | *YES* | |
| *Tick spp 16S rRNA* | *YES* | *YES* [2] | |
| *Amblyomma variegatum ITS2* | *YES* | *YES* | |
| *Rhipicephalus microplus ITS2* | *YES* | *YES* | |
| *Rhipicephalus sanguineus s.l. ITS2* | *YES* | *YES* | |

[1] Designs removed from the screening analysis.

[2] Tick spp. primers required to be part of the pre-amplification mix in order to achieve correct detection signals.

Regarding the seven designs displaying unexpected signals, we decided to explore the hypothesis of potential co-infection in controls corresponding to DNA from ticks or blood samples using conventional PCR and amplicon sequencing (Table A1).

Both the *Rickettsia* spp. and *Rickettsia felis* designs gave positive results when testing the *Borrelia lonestari*, *Panola Mountain Ehrlichia*, *Ehrlichia ewingii* and *Ehrlichia chaffensis* controls. All these samples corresponded to DNA extracted from infected *Amblyomma americanum* collected in the field (USA). This result supports the fact that a *Rickettsia* species may have been present in these samples, whether it was *Rickettsia felis* or a closely related *Rickettsia* spp. infecting *Amblyomma americanum* ticks. When

testing the Panola Mountain Ehrlichia sample for *Rickettsia* spp. by conventional PCR targeting the gltA gene [65], we obtained a 382 bp sequence (accession number MK049843) sharing 99% sequence identity with *Rickettsia* spp. (MF511253.1) (Table A2). This result, in addition to natural co-infections documented in *Amblyomma americanum* ticks, suggest that the detection of *Rickettsia* spp. – within the four outgroup positive controls corresponding to *Amblyomma americanum* DNA – in this assay did not correspond to cross-reactions [73,74]. Moreover, the *Amblyomma variegatum* sample, corresponding to ticks collected from the field, was found to be positive for both *Rickettsia* spp. and *Rickettsia africae*. As this sample originated from Guadeloupe, where *Rickettsia africae* circulates, this result may have been in agreement with a natural infection [9].

**Table A2.** Taxonomic assignment of the sequences obtained after sequencing PCR products to confirm the presence of co-infections in complex control samples corresponding to DNA extracted from wild ticks or blood samples. AN: accession number, % I: percentage identity, % C: percentage coverage.

| Tested control sample | Tested for | AN | Length (bp) | Closest homology | % I | % C | AN |
|---|---|---|---|---|---|---|---|
| *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) | *Rickettsia* spp. | MK049843 | 382 | *Rickettsia* spp. | 99 | 100 | MF511253.1 |
| *Neoehrlichia mikurensis* (Infected rodent blood) | *Hepatozoon* spp. | MK071735 | 169 | *Hepatozoon* spp. | 99 | 100 | AB771515.1 |
| *Anaplasma marginale* (Infected cow blood) | Apicomplexa | MK071737 | 104 | *Babesia spp., Theileria* spp. (including *B. bigemina*) | 100 | 99 | MG604302.1 |
| *Panola mountain Ehrlichia* (Infected *A. americanum* ticks) | Apicomplexa | MK071736 | 102 | *Theileria* spp. (including *T. cervi*) | 98 | 100 | MH085203.1 |
| *Borrelia lonestari* (Infected *A. americanum* ticks) | Apicomplexa | | | | | | |

Moreover, the *Rhipicephalus sanguineus* s.l. sample was positive with four designs targeting parasites – Apicomplexa, *Hepatozoon* spp., *Hepatozoon canis* and *Hepatozoon americanum*. Since *Rhipicephalus sanguineus* s.l. ticks are involved in the epidemiology of tick-borne parasites including *Hepatozoon* spp., these results strongly suggest the occurrence of such parasites in the biological sample [75]. Thus, these observations suggest that our designs were actually capable of detecting pathogens present in naturally infected ticks rather than giving an unsuspected cross-reaction. The *Neoehrlichia mikurensis* sample, corresponding to DNA extracted from rodent blood, was also found to be positive for both Apicomplexa and *Hepatozoon* spp. The amplicon obtained from this sample with the *Hepatozoon* spp. design was sequenced. The obtained 169 bp sequence (accession number MK071735) displayed 99% sequence identity with the *Hepatozoon* spp. sequences (AB771515.1) (Table A2). As rodents can be infected with *Hepatozoon* parasites, this result could also reflect a natural infection [76]. The *Anaplasma marginale* sample, corresponding to a blood sample from an experimentally infected cow, was also found to be positive for both Apicomplexa and Babesia bigemina. The amplicon obtained from this sample with the Apicomplexa design was sequenced. The obtained 104 bp sequence (accession number MK071737) displayed 99% sequence identity with the Apicomplexan sequences, including *Babesia bigemina* (MG604302.1) (Table A2). As *Babesia bigemina* and *Anaplasma marginale* are two cattle pathogens that have often evolved in the same region and are transmitted by the same vector tick, co-infections with these two pathogens have already been reported [77]. Thus, this cross-reaction may again have been a reflection of parasite co-infections. Lastly, Apicomplexa also gave positive results in the *Borrelia lonestari* and *Panola Mountain Ehrlichia* controls. As highlighted previously, these two controls corresponded to DNA extracted from *A. americanum* ticks. The amplicons obtained from these two samples with the Apicomplexa design were sequenced. The two obtained 102 bp sequences (accession number MK071736) were identical and

displayed 98% sequence identity with the *Theileria cervi* sequences (MH085203.1) (Table A2). As *Theileria cervi* is a common deer pathogen found in *Amblyomma americanum*, the occurrence of this parasite in these two control samples could explain the unexpected signals [78,79].

## Appendix B

Analysis of the co-infections detected in *Amblyomma variegatum* and *Rhipicephalus microplus* ticks collected in Guadeloupe and Martinique.

Here we reported the co-infections observed in *Amblyomma variegatum* (n=132 samples), *Rhipicephalus microplus* collected in Guadeloupe (n=116 samples, including individual and pooled specimens) and Martinique (n= 275 samples, including individual and pooled specimens) (Table A3). We combined the results obtained with the *Rickettsia* spp. and the *R. africae* design, assuming that only *R. africae* have been detected in the Caribbean samples analyzed here. In addition, *Anaplasma* spp. results here correspond to sample only positive for *Anaplasma* spp. and not for *A. marginale*, as well, *Ehrlichia* spp. results here correspond to sample only positive for *Ehrlichia* spp. and not for *E. ruminantium*.

**Table A3.** Co-infection analysis in *A. variegatum* (n=132 samples) and *R. microplus* collected in Guadeloupe (n=116 samples) and Martinique (n=275 samples).

| | | ***A. variegatum*** **sample (percentage)** | ***R. microplus*** **sample from Guadeloupe (percentage)** | ***R. microplus*** **sample from Martinique (percentage)** |
|---|---|---|---|---|
| | Total of sample | 132 | 116 | 275 |
| | Total of non-infected sample | 1 (0.8%) | 51 (44%) | 52 (19%) |
| | Total of infected sample | 131 (99.2%) | 65 (56%) | 223 (81%) |
| Single infections | Total | 69 (52.3%) | 47 (40.5%) | 99 (36%) |
| | *R. africae* [1] | 68 (51.5%) | 15 (12.9%) | 0 |
| | *Anaplasma* spp. [2] | | 1 (0.9%) | 10 (3.6%) |
| | *A. marginale* | 0 | 2 (1.7%) | 22 (8%) |
| | *Ehrlichia* spp. [3] | 0 | 3 (2.6%) | 43 (15.6%) |
| | *Borrelia* spp. | 0 | 0 | 3 (1.1%) |
| | *B. bigemina* | 0 | 1 (0.9%) | 7 (2.5%) |
| | *T. velifera* | 1 (0.8%) | 25 (21.6%) | 14 (5.1%) |
| Co-infections (2) | *Total* | 53 (40.2%) | 16 (13.8%) | 88 (32%) |
| | *R. africae* [1] / *T. velifera* | 47 (35.6%) | 6 (5.2%) | 0 |
| | *R. africae* [1] / *Borrelia* spp. | 3 (2.3%) | 0 | 0 |
| | *R. africae* [1]/ *E. ruminantium* | 3 (2.3%) | 0 | 0 |
| | *Anaplasma* spp. [2] / *Borrelia* spp. | 0 | 1 (0.9%) | 0 |
| | *Anaplasma* spp. [2] / *T. velifera* | 0 | 2 (1.7%) | 3 (1.1%) |
| | *Anaplasma* spp. [2] / *Ehrlichia* spp. [3] | 0 | 0 | 6 (2.2%) |
| | *A. marginale* / *T. velifera* | 0 | 2 (1.7%) | 10 (3.6%) |
| | *A. marginale* / *Borrelia* spp. | 0 | 0 | 3 (1.1%) |
| | *A. marginale* / *Ehrlichia* spp. [3] | 0 | 0 | 39 (14.29%) |
| | *A. marginale* / *B. bigemina* | 0 | 0 | 4 (1.5%) |

| | | | | |
|---|---|---|---|---|
| | *Ehrlichia* spp. [3] / *R. africae* [1] | 0 | 2 (1.7%) | 0 |
| | *Ehrlichia* spp. [3] / *T. velifera* | 0 | 1 (0.9%) | 8 (2.9%) |
| | *Ehrlichia* spp. [3] / *Borrelia* spp. | 0 | 0 | 3 (1.1%) |
| | *Ehrlichia* spp. [3] / *Leishmania* spp. | 0 | 0 | 1 (0.4%) |
| | *T. velifera* / *B. bigemina* | 0 | 0 | 10 (3.6%) |
| | *T. velifera* / *T. mutans* | 0 | 2 (1.7%) | 1 (0.4%) |
| Co-infections (3) | *Total* | 8 (6.1%) | 2 (1.7%) | 26 (9.5%) |
| | *R.africae* [1] / *E. ruminantium* / *T. velifera* | 3 (2.3%) | 0 | 0 |
| | *R. africae* [1] / *Borrelia spp.* / *T. velifera* | 3 (2.3%) | 0 | 0 |
| | *R. africae* [1] / *T. velifera* / *T. mutans* | 2 (1.5%) | 1 (0.6%) | 0 |
| | *R. africae* [1] / *A. marginale* / *Ehrlichia* spp. [3] | 0 | 1 (0.6%) | 0 |
| | *A. marginale* / *Ehrlichia* spp. [3] / *B. bigemina* | 0 | 0 | 8 (2.9%) |
| | *A. marginale* / *Ehrlichia* spp. [3] / *B.bovis* | 0 | 0 | 1 (0.4%) |
| | *A. marginale* / *Ehrlichia* spp. [3] / *T. velifera* | 0 | 0 | 15 (5.5%) |
| | *Anaplasma* spp. [2] / *Ehrlichia* spp. [3] / *B. bigemina* | 0 | 0 | 1 (0.4%) |
| | *Anaplasma* spp. [2] / *Ehrlichia* spp. [3] / *T.velifera* | 0 | 0 | 1 (0.4%) |
| Co-infections (4) | *Total* | 1 (0.8%) | 0 | 9 (3.3%) |
| | *R. africae* [1] / *T. velifera* / *Borrelia* spp. / *E. ruminantium* | 1 (0.8%) | 0 | 0 |
| | *A. marginale* / *Borrelia* spp. / *T. velifera* / *T. mutans* | 0 | 0 | 1 (0.4%) |
| | *A. marginale* / *Ehrlichia* spp. [3] / *T. velifera* / *B. bigemina* | 0 | 0 | 5 (1.8%) |
| | *A. marginale* / *Ehrlichia* spp. [3] / *T. velifera* / *B.bovis* | 0 | 0 | 1 (0.4%) |
| | *A. marginale* / *Leishmania* spp. / *T. velifera* / *T. mutans* | 0 | 0 | 1 (0.4%) |
| | *Anaplasma* spp. [2] / *Ehrlichia* spp. [3] / *Borrelia* spp. / *T. velifera* | 0 | 0 | 1 (0.4%) |
| Co-infections (5) | *Total* | 0 | 0 | 1 (0.4%) |
| | *A. marginale* / *Ehrlichia* spp. [3] / *Borrelia* spp. / *T. velifera* / *T. mutans* | 0 | 0 | 1 (0.4%) |

[1] Assuming that all the *Rickettsia* spp. found in *Rhipicephalus microplus* samples from Guadeloupe were *R. africae*.
[2] Sample positive for *Anaplasma* spp. and not for *A. marginale*.
[3] Sample positive for *Ehrlichia* spp. and not for *E. ruminantium*.

Almost all of the *A. variegatum* samples from Guadeloupe were infected with at least one pathogen (99.2%) (Table A3). Interestingly, only 56% of the *R. microplus* samples from Guadeloupe were infected with at least one pathogen, whereas this rate reach 81% of the *R. microplus* from Martinique (Table A3). Most of the positive samples corresponded to single infection or co-infection with two pathogens in both tick species. Then, less than 10% of the tick samples displayed co-infections with three pathogens (Table A3). Finally, only one *A. variegatum* sample from Guadeloupe

and *nine R. microplus* samples from Martinique were co-infected with four pathogens and one *R. microplus* from Martinique was found infected with five pathogens (Table A3).

The majority of the *A. variegatum* samples displayed single infections with *R. africae* (52%) or co-infections with *R. africae* and *T. velifera* (36%) (Table A3). Nevertheless, no negative or positive association have been detected between the microorganisms detected in *A. variegatum* samples (Table A4). At least, the presence of *R. africae* do not seem to interfere with the presence of *T. velifera* (Table A4).

**Table A4.** Co-occurrence matrix of the microorganisms detected in *A. variegatum* samples collected in Guadeloupe (n=132 samples). No negative or positive association have been detected when performing a co-occurrence test using the co-occur function ($\alpha$=0.05) and R version 3.6.0 (2019-04-26).

| | *E. ruminantium* | *Borrelia* spp. | *R. africae* | *T. mutans* | *T. velifera* |
|---|---|---|---|---|---|
| *E. ruminantium* | 7 | **1** | **7** | **0** | **4** |
| *Borrelia* spp. | | 7 | **7** | **0** | **4** |
| *R. africae* | | | 130 | **2** | **56** |
| *T. mutans* | | | | 2 | **2** |
| *T. velifera* | | | | | 57 |

Among the 116 *R. microplus* samples from Guadeloupe analyzed here, most of the positive samples presented single-infection (40.5%), with *R. africae* (12.9%) or *T. velifera* (21.6%) (Table A3). As *R. microplus* is not considered as a vector of both of these microorganisms, we made the hypothesis of a possible contamination of this tick species via infected bovine blood still present in engorged tick, and/or via co-feeding with infected *A. variegatum* ticks. Interestingly, if we remove *R. africae*, *T. velifera* and *T. mutans* from the screening analysis, the percentage of infected *R. microplus* from Guadeloupe dropped drastically to 13.8% (16/116 samples infected with at least one pathogen). This observation is particularly surprising when comparing this rate to the 81% infected *R. microplus* from Martinique (see below). When testing for co-occurrence linkage, two positive associations have been detected between the microorganisms detected in *R. microplus* samples from Guadeloupe, including *T. velifera* / *T. mutans*, and *Anaplasma* spp. / *Borrelia* spp. (Table A5). Co-infections and positive association between *T. velifera* and *T. mutans* have already been reported in the literature, such as in cattle sera from Uganda, and Kenya [80,81]. Regarding the few samples positive for *Anaplasma* spp. and *Borrelia* spp., the result of the co-occurrence test should be taken with cautions and deserve further investigation. Nevertheless, no exclusion seemed to occur between the pathogens/microorganisms detected in *R. microplus* from Guadeloupe (Table A5).

**Table A5.** Co-occurrence matrix of the microorganisms detected in *R. microplus* samples collected in Guadeloupe (n=116 samples). *: Positive association detected when performing a co-occurrence test using the co-occur function ($\alpha$=0.05) and R version 3.6.0 (2019-04-26).

| | ***A. marginale*** | ***Borrelia* spp.** | ***R. africae*** | ***T. mutans*** | ***T. velifera*** | ***B. bigemina*** | ***Anaplasma* spp.[1]** | ***Ehrlichia* spp.[2]** |
|---|---|---|---|---|---|---|---|---|
| *A. marginale* | 5 | 0 | 1 | 0 | 2 | 0 | 0 | 1 |
| *Borrelia* spp. | | 1 | 0 | 0 | 0 | 0 | **1*** | 0 |
| *R. africae* | | | 25 | 1 | 7 | 0 | 0 | 3 |
| *T. mutans* | | | | 3 | **3*** | 0 | 0 | 0 |
| *T. velifera* | | | | | 39 | 0 | 2 | 1 |
| *B. bigemina* | | | | | | 1 | 0 | 0 |
| *Anaplasma* spp.[1] | | | | | | | 4 | 0 |
| *Ehrlichia* spp.[2] | | | | | | | | 7 |

[1] Sample positive for *Anaplasma* spp. and not for *A. marginale*.

[2] Sample positive for *Ehrlichia* spp. and not for *E. ruminantium*.

Among the 275 *R. microplus* samples from Guadeloupe, most of the sample presented single-infection (36%) and co-infections with two pathogens (32%) (Table A1). Five positive associations have been detected between the microorganisms detected in *R. microplus* samples from Martinique (Table A6). *T. mutans* have been found in positive association with *T. velifera*, as previously observed in the same tick species from Guadeloupe, and with *Leishmania* spp. and *Borrelia* spp. In addition, *T. velifera* were found in positive association with *B. bigemina* and *Ehrlichia spp.* with *A. marginale.* Finally, no negative association have been reported between the pathogens/microorganisms detected in *R. microplus* from Martinique (Table A6).

**Table A6.** Co-occurrence matrix of the microorganisms detected in *R. microplus* samples collected in Martinique (n=275 samples). (*) Positive associations detected when performing a co-occurrence test using the co-occur function ($\alpha$=0.05) and R version 3.6.0 (2019-04-26).

| | ***A. marginale*** | ***Borrelia* spp.** | ***Leishmania* spp.** | ***T. mutans*** | ***T. velifera*** | ***B. bigemina*** | ***B. bovis*** | ***Anaplasma* spp.[1]** | ***Ehrlichia* spp.[2]** |
|---|---|---|---|---|---|---|---|---|---|
| *A. marginale* | 111 | 5 | 1 | 3 | 34 | 17 | 2 | 0 | **70*** |
| *Borrelia* spp. | | 12 | 0 | **2*** | 3 | 0 | 0 | 1 | 5 |
| *Leishmania* spp. | | | 2 | **1*** | 1 | 0 | 0 | 0 | 1 |
| *T. mutans* | | | | 4 | **4*** | 0 | 0 | 0 | 1 |
| *T. velifera* | | | | | 72 | **15*** | 1 | 5 | 32 |
| *B. bigemina* | | | | | | 35 | 0 | 1 | 14 |
| *B. bovis* | | | | | | | 2 | 0 | 2 |
| *Anaplasma* spp.1 | | | | | | | | 22 | 9 |
| *Ehrlichia* spp.2 | | | | | | | | | 134 |

[1] Sample positive for *Anaplasma* spp. and not for *A. marginale*.
[2] Sample positive for *Ehrlichia* spp. and not for *E. ruminantium*.

## References


1. Dantas-Torres, F.; Chomel, B.B.; Otranto, D. Ticks and tick-borne diseases: a One Health perspective. *Trends Parasitol.* **2012**, *28*, 437–446. Doi: 10.1016/j.pt.2012.07.003
2. George, J.E.; Davey, R.B.; Pound, J.M. Introduced ticks and tick-borne diseases: the threat and approaches to eradication. *Vet. Clin. North Am. Food Anim. Pract.* **2002**, *18*, 401–416, vi.
3. Gondard, M.; Cabezas-Cruz, A.; Charles, R.A.; Vayssier-Taussat, M.; Albina, E.; Moutailler, S. Ticks and Tick-Borne Pathogens of the Caribbean: Current Understanding and Future Directions for More Comprehensive Surveillance. *Front Cell Infect Microbiol* **2017**, *7*, 490. Doi:10.3389/fcimb.2017.00490.
4. Barré, N.; Uilenberg, G. Spread of parasites transported with their hosts: case study of two species of cattle tick. *Rev. - Off. Int. Epizoot.* **2010**, *29*, 149–160, 135–147.
5. Morel, P.-C. Etudes sur les tiques du bétail en Guadeloupe et Martinique. II. Agents pathogènes transmis par les tiques. *Revue d'élevage et de médecine vétérinaire des pays tropicaux* **1967**, *20*, 291–299. Doi:10.19182/remvt.7508.
6. Uilenberg, G.; Camus, E.; Barré, N. [Existence of Theileria mutans and Theileria velifera (Sporozoa, Theileriidae) in Guadeloupe (French West Indies)]. *Rev Elev Med Vet Pays Trop* **1983**, *36*, 261–264.
7. Camus, E.; Barre, N. Vector situation of tick-borne diseases in the Caribbean islands. *Veterinary Parasitology* **1995**, *57*, 167–176. Doi: 10.1016/0304-4017(94)03118-G.
8. Alonso, M.; Arellano-Sota, C.; Cereser, V.H.; Cordoves, C.O.; Guglielmone, A.A.; Kessler, R.; Mangold,

A.J.; Nari, A.; Patarroyo, J.H.; Solari, M.A. Epidemiology of bovine anaplasmosis and babesiosis in Latin America and the Caribbean. *Rev. - Off. Int. Epizoot.* **1992**, *11*, 713–733.

9. Parola, P.; Vestris, G.; Martinez, D.; Brochier, B.; Roux, V.; Raoult, D. Tick-borne rickettiosis in Guadeloupe, the French West Indies: isolation of *Rickettsia africae* from *Amblyomma variegatum* ticks and serosurvey in humans, cattle, and goats. *Am. J. Trop. Med. Hyg.* **1999**, *60*, 888–893.
10. Camus, E.; Montenegro-James, S. Bovine anaplasmosis and babesiosis in the Lesser Antilles: risk assessment of an unstable epidemiologic situation. *Vet. Res.* **1994**, *25*, 313–317.
11. Barre, N.; Garris, G.I.; Borel, G.; Camus, E. Hosts and Population Dynamics of *Amblyomma variegatum* (Acari: Ixodidae) on Guadeloupe, French West Indies. *J Med Entomol* **1988**, *25*, 111–115. Doi: 10.1093/jmedent/25.2.111.
12. Vachiéry, N.; Jeffery, H.; Pegram, R.; Aprelon, R.; Pinarello, V.; Kandassamy, R.L.Y.; Raliniaina, M.; Molia, S.; Savage, H.; Alexander, R.; et al. *Amblyomma variegatum* ticks and heartwater on three Caribbean Islands. *Ann. N. Y. Acad. Sci.* **2008**, *1149*, 191–195. Doi: 10.1196/annals.1428.081.
13. Parola, P.; Attali, J.; Raoult, D. First detection of *Rickettsia africae* on Martinique, in the French West Indies. *Ann Trop Med Parasitol* **2003**, *97*, 535–537. Doi: 10.1179/000349803225001382.
14. Kelly, P.; Lucas, H.; Beati, L.; Yowell, C.; Mahan, S.; Dame, J. *Rickettsia africae* in *Amblyomma variegatum* and domestic ruminants on eight Caribbean islands. *J. Parasitol.* **2010**, *96*, 1086–1088. Doi: 10.1645/GE-2552.1.
15. Raoult, D.; Fournier, P.E.; Fenollar, F.; Jensenius, M.; Prioe, T.; de Pina, J.J.; Caruso, G.; Jones, N.; Laferl, H.; Rosenblatt, J.E.; et al. *Rickettsia africae*, a Tick-Borne Pathogen in Travelers to Sub-Saharan Africa. *New England Journal of Medicine* **2001**, *344*, 1504–1510. Doi: 10.1056/NEJM200105173442003.
16. Estrada-Peña, A.; Gray, J.S.; Kahl, O.; Lane, R.S.; Nijhof, A.M. Research on the ecology of ticks and tick-borne pathogens—methodological principles and caveats. *Front Cell Infect Microbiol* **2013**, *3*. Doi: 10.3389/fcimb.2013.00029.
17. Cabezas-Cruz, A.; Vayssier-Taussat, M.; Greub, G. Tick-borne pathogen detection: what's new? *Microbes Infect.* **2018**. Doi: 10.1016/j.micinf.2017.12.015.
18. Michelet, L.; Delannoy, S.; Devillers, E.; Umhang, G.; Aspan, A.; Juremalm, M.; Chirico, J.; van der Wal, F.J.; Sprong, H.; Boye Pihl, T.P.; et al. High-throughput screening of tick-borne pathogens in Europe. *Front Cell Infect Microbiol* **2014**, *4*. Doi: 10.3389/fcimb.2014.00103.
19. Dupraz, M.; Toty, C.; Devillers, E.; Blanchon, T.; Elguero, E.; Vittecoq, M.; Moutailler, S.; McCoy, K.D. Population structure of the soft tick *Ornithodoros maritimus* and its associated infectious agents within a colony of its seabird host *Larus michahellis*. *Int J Parasitol Parasites Wildl* **2017**, *6*, 122–130. Doi: 10.1016/j.ijppaw.2017.05.001.
20. Gioia, G.V.; Vinueza, R.L.; Marsot, M.; Devillers, E.; Cruz, M.; Petit, E.; Boulouis, H.J.; Moutailler, S.; Monroy, F.; Coello, M.A.; et al. Bovine anaplasmosis and tick-borne pathogens in cattle of the Galapagos Islands. *Transbound Emerg Dis* **2018**, *65*, 1262–1271. Doi: 10.1111/tbed.12866.
21. Gondard, M.; Michelet, L.; Nisavanh, A.; Devillers, E.; Delannoy, S.; Fach, P.; Aspan, A.; Ullman, K.; Chirico, J.; Hoffmann, B.; et al. Prevalence of tick-borne viruses in *Ixodes ricinus* assessed by high-throughput real-time PCR. *Pathog Dis* **2018**, *76*. Doi: 10.1093/femspd/fty083.
22. Malmsten, J.; Dalin, A.-M.; Moutailler, S.; Devillers, E.; Gondard, M.; Felton, A. Vector-Borne Zoonotic Pathogens in Eurasian Moose (Alces alces alces). *Vector Borne Zoonotic Dis.* **2018**. Doi: 10.1089/vbz.2018.2277.
23. Ahantarig, A.; Trinachartvanit, W.; Baimai, V.; Grubhoffer, L. Hard ticks and their bacterial

endosymbionts (or would be pathogens). *Folia Microbiol. (Praha)* **2013**, *58*, 419–428. Doi: 10.1007/s12223-013-0222-1.

24. Duron, O.; Binetruy, F.; Noël, V.; Cremaschi, J.; McCoy, K.D.; Arnathau, C.; Plantard, O.; Goolsby, J.; Pérez de León, A.A.; Heylen, D.J.A.; et al. Evolutionary changes in symbiont community structure in ticks. *Mol. Ecol.* **2017**, *26*, 2905–2921. Doi: 10.1111/mec.14094.
25. Bonnet, S.; Jouglin, M.; Malandrin, L.; Becker, C.; Agoulon, A.; L'hostis, M.; Chauvin, A. Transstadial and transovarial persistence *of Babesia divergens* DNA in *Ixodes ricinus* ticks fed on infected blood in a new skin-feeding technique. *Parasitology* **2007**, *134*, 197–207. Doi: 10.1017/S0031182006001545.
26. Howell, J.M.; Ueti, M.W.; Palmer, G.H.; Scoles, G.A.; Knowles, D.P. Transovarial Transmission Efficiency of *Babesia bovis* Tick Stages Acquired by *Rhipicephalus* (*Boophilus*) *microplus* during Acute Infection. *J Clin Microbiol* **2007**, *45*, 426–431. Doi: 10.1128/JCM.01757-06.
27. Chauvin, A.; Moreau, E.; Bonnet, S.; Plantard, O.; Malandrin, L. *Babesia* and its hosts: adaptation to long-lasting interactions as a way to achieve efficient transmission. *Vet. Res.* **2009**, *40*, 37. Doi: 10.1051/vetres/2009020.
28. Robinson, J.B.; Eremeeva, M.E.; Olson, P.E.; Thornton, S.A.; Medina, M.J.; Sumner, J.W.; Daschi, G.A. New approaches to detection and identification of *Rickettsia africae* and *Ehrlichia ruminantium* in *Amblyomma variegatum* (Acari: Ixodidae) ticks from the Caribbean. *J. Med. Entomol.* **2009**, *46*, 942–951.
29. Socolovschi, C.; Huynh, T.P.; Davoust, B.; Gomez, J.; Raoult, D.; Parola, P. Transovarial and trans-stadial transmission of *Rickettsiae africae* in *Amblyomma variegatum* ticks. *Clin. Microbiol. Infect.* **2009**, *15 Suppl 2*, 317–318. Doi: 10.1111/j.1469-0691.2008.02278.x.
30. Maina, A.N.; Jiang, J.; Omulo, S.A.; Cutler, S.J.; Ade, F.; Ogola, E.; Feikin, D.R.; Njenga, M.K.; Cleaveland, S.; Mpoke, S.; et al. High Prevalence of *Rickettsia africae* Variants in *Amblyomma variegatum* Ticks from Domestic Mammals in Rural Western Kenya: Implications for Human Health. *Vector Borne Zoonotic Dis* **2014**, *14*, 693–702. Doi: 10.1089/vbz.2014.1578.
31. Yssouf, A.; Socolovschi, C.; Kernif, T.; Temmam, S.; Lagadec, E.; Tortosa, P.; Parola, P. First molecular detection of *Rickettsia africae* in ticks from the Union of the Comoros. *Parasites & Vectors* **2014**, *7*, 444. Doi: 10.1186/1756-3305-7-444.
32. Matsimbe, A.M.; Magaia, V.; Sanches, G.S.; Neves, L.; Noormahomed, E.; Antunes, S.; Domingos, A. Molecular detection of pathogens in ticks infesting cattle in Nampula province, Mozambique. *Exp Appl Acarol* **2017**, *73*, 91–102. Doi: 10.1007/s10493-017-0155-5.
33. Rodriguez O.N.; Espaine, L.; Rivas, A.; Rodriguez, P.. Epidemiology of cattle diseases caused by haemoparasites in Cuba. Available online: http://www.cabi.org/isc/abstract/19902200923 (accessed on Mar 25, 2017).
34. Woolhouse, M.E.J.; Thumbi, S.M.; Jennings, A.; Chase-Topping, M.; Callaby, R.; Kiara, H.; Oosthuizen, M.C.; Mbole-Kariuki, M.N.; Conradie, I.; Handel, I.G.; et al. Co-infections determine patterns of mortality in a population exposed to parasite infection. *Science Advances* **2015**, *1*, e1400026. Doi: 10.1126/sciadv.1400026.
35. Parola, P.; Inokuma, H.; Camicas, J.L.; Brouqui, P.; Raoult, D. Detection and identification of spotted fever group Rickettsiae and Ehrlichiae in African ticks. *Emerging Infect. Dis.* **2001**, *7*, 1014–1017. Doi: 10.3201/eid0706.010616.
36. Inokuma, H.; Beppu, T.; Okuda, M.; Shimada, Y.; Sakata, Y. Detection of ehrlichial DNA in *Haemaphysalis* ticks recovered from dogs in Japan that is closely related to a novel *Ehrlichia* sp. found in cattle ticks from Tibet, Thailand, and Africa. *J. Clin. Microbiol.* **2004**, *42*, 1353–1355.

37. Loftis, A.D.; Reeves, W.K.; Szumlas, D.E.; Abbassy, M.M.; Helmy, I.M.; Moriarity, J.R.; Dasch, G.A. Rickettsial agents in Egyptian ticks collected from domestic animals. *Exp. Appl. Acarol.* **2006**, *40*, 67–81. Doi: 10.1007/s10493-006-9025-2.
38. Kang, Y.-J.; Diao, X.-N.; Zhao, G.-Y.; Chen, M.-H.; Xiong, Y.; Shi, M.; Fu, W.-M.; Guo, Y.-J.; Pan, B.; Chen, X.-P.; et al. Extensive diversity of Rickettsiales bacteria in two species of ticks from China and the evolution of the Rickettsiales. *BMC Evolutionary Biology* **2014**, *14*, 167. Doi: 10.1186/s12862-014-0167-2.
39. Lu, M.; Tian, J.-H.; Yu, B.; Guo, W.-P.; Holmes, E.C.; Zhang, Y.-Z. Extensive diversity of rickettsiales bacteria in ticks from Wuhan, China. *Ticks Tick Borne Dis* **2017**, *8*, 574–580. Doi: 10.1016/j.ttbdis.2017.03.006.
40. Guo, W.-P.; Tian, J.-H.; Lin, X.-D.; Ni, X.-B.; Chen, X.-P.; Liao, Y.; Yang, S.-Y.; Dumler, J.S.; Holmes, E.C.; Zhang, Y.-Z. Extensive genetic diversity of Rickettsiales bacteria in multiple mosquito species. *Sci Rep* **2016**, *6*, 38770. Doi: 10.1038/srep38770.
41. Yparraguirre, L.A.; Machado-Ferreira, E.; Ullmann, A.J.; Piesman, J.; Zeidner, N.S.; Soares, C.A.G. A hard tick relapsing fever group spirochete in a Brazilian *Rhipicephalus* (*Boophilus*) *microplus*. *Vector Borne Zoonotic Dis.* **2007**, *7*, 717–721. Doi: 10.1089/vbz.2007.0144.
42. Hagen, R.M.; Frickmann, H.; Ehlers, J.; Krüger, A.; Margos, G.; Hizo-Teufel, C.; Fingerle, V.; Rakotozandrindrainy, R.; Kalckreuth, V. von; Im, J.; et al. Presence of *Borrelia* spp. DNA in ticks, but absence of *Borrelia* spp. and of *Leptospira* spp. DNA in blood of fever patients in Madagascar. *Acta Trop.* **2018**, *177*, 127–134. Doi: 10.1016/j.actatropica.2017.10.002.
43. Kumsa, B.; Socolovschi, C.; Raoult, D.; Parola, P. New Borrelia species detected in ixodid ticks in Oromia, Ethiopia. *Ticks Tick Borne Dis* **2015**, *6*, 401–407. Doi: 10.1016/j.ttbdis.2015.03.009.
44. Ehounoud, C.B.; Yao, K.P.; Dahmani, M.; Achi, Y.L.; Amanzougaghene, N.; Kacou N'Douba, A.; N'Guessan, J.D.; Raoult, D.; Fenollar, F.; Mediannikov, O. Multiple Pathogens Including Potential New Species in Tick Vectors in Côte d'Ivoire. *PLoS Negl Trop Dis* **2016**, *10*. Doi: 10.1371/journal.pntd.0004367.
45. Estrada-Peña, A.; Álvarez-Jarreta, J.; Cabezas-Cruz, A. Reservoir and vector evolutionary pressures shaped the adaptation of *Borrelia*. *Infection, Genetics and Evolution* **2018**. Doi: 10.1016/j.meegid.2018.03.023.
46. Rodríguez, I.; Fernández, C.; Cinco, M.; Pedroso, R.; Fuentes, O. Do Antiborrelial Antibodies Suggest Lyme Disease in Cuba? *Emerg Infect Dis* **2004**, *10*, 1698–1700. Doi: 10.3201/eid1009.031048.
47. Rodríguez, I.; Fernández, C.; Sánchez, L.; Martínez, B.; Siegrist, H.H.; Lienhard, R. Prevalence of antibodies to *Borrelia burgdorferi* sensu stricto in humans from a Cuban village. *Braz J Infect Dis* **2012**, *16*, 82–85. Doi: http://dx.doi.org/10.1590/S1413-86702012000100015.
48. Dessau, R.B. Infection due to *Borrelia burgdorferi* most likely does not occur in Cuba. *Braz J Infect Dis* **2012**, *16*, 404; author reply 405-406. Doi: 10.1016/j.bjid.2012.06.007.
49. Flanigan, T.P.; Schwan, T.G.; Armstrong, C.; Voris, V.; P, L.; Salata, R.A. Relapsing Fever in the US Virgin Islands: A Previously Unrecognized Focus of Infection. *J Infect Dis* **1991**, *163*, 1391–1392. Doi: 10.1093/infdis/163.6.1391.
50. Sharma, A.; Jaimungal, S.; Basdeo-Maharaj, K.; Rao, A.V.C.; Teelucksingh, S. Erythema Migrans–like Illness among Caribbean Islanders. *Emerg Infect Dis* **2010**, *16*, 1615–1617. Doi: 10.3201/eid1610.100587.
51. Pothirat, T.; Tantiworawit, A.; Chaiwarith, R.; Jariyapan, N.; Wannasan, A.; Siriyasatien, P.; Supparatpinyo, K.; Bates, M.D.; Kwakye-Nuako, G.; Bates, P.A. First Isolation of *Leishmania* from Northern Thailand: Case Report, Identification as *Leishmania martiniquensis* and Phylogenetic Position within the *Leishmania enriettii* Complex. *PLoS Negl Trop Dis* **2014**, *8*. Doi: 10.1371/journal.pntd.0003339.
52. Espinosa, O.A.; Serrano, M.G.; Camargo, E.P.; Teixeira, M.M.G.; Shaw, J.J. An appraisal of the taxonomy

and nomenclature of trypanosomatids presently classified as *Leishmania* and Endotrypanum. *Parasitology* **2018**, *145*, 430–442. Doi: 10.1017/S0031182016002092.

53. Leelayoova, S.; Siripattanapipong, S.; Manomat, J.; Piyaraj, P.; Tan-ariya, P.; Bualert, L.; Mungthin, M. Leishmaniasis in Thailand: A Review of Causative Agents and Situations. *Am J Trop Med Hyg* **2017**, *96*, 534–542. Doi: 10.4269/ajtmh.16-0604.
54. Steverding, D. The history of leishmaniasis. *Parasit Vectors* **2017**, *10*, 82. Doi : 10.1186/s13071-017-2028-5.
55. Desbois, N.; Pratlong, F.; Quist, D.; Dedet, J.-P. *Leishmania* (*Leishmania*) *martiniquensis* n. sp. (Kinetoplastida: Trypanosomatidae), description of the parasite responsible for cutaneous leishmaniasis in Martinique Island (French West Indies). *Parasite* **2014**, *21*, 12. Doi: 10.1051/parasite/2014011.
56. Liautaud, B.; Vignier, N.; Miossec, C.; Plumelle, Y.; Kone, M.; Delta, D.; Ravel, C.; Cabié, A.; Desbois, N. First case of visceral leishmaniasis caused by *Leishmania martiniquensis*. *Am. J. Trop. Med. Hyg.* **2015**, *92*, 317–319. Doi: 10.4269/ajtmh.14-0205.
57. Dantas-Torres, F. Ticks as vectors of *Leishmania* parasites. *Trends Parasitol.* **2011**, *27*, 155–159. Doi: 10.1016/j.pt.2010.12.006.
58. Rakhshanpour, A.; Malmasi, A.; Mohebali, M.; Nabian, S.; Mirhendi, H.; Zarei, Z.; Dalimi, A.; Mohammadiha, A.; Akhoundi, B.; Azarm, A. Transmission of *Leishmania infantum* by *Rhipicephalus sanguineus* (Acari: Ixodidae) in Dogs. *Iran J Parasitol* **2017**, *12*, 482–489.
59. Rojas-Jaimes, J.E.; Correa-Nuñez, G.H.; Rojas, N.; Cáceres-Rey, O. Detection of *Leishmania* (*V*) *guyanensis* in *Rhipicephalus* (*Boophilus*) *microplus* (Acari: Ixodidae) collected from *Pecari tajacu*. *Biomedica* **2017**, *37*, 208–214.
60. Alam, M.Z.; Rahman, M.M.; Akter, S.; Talukder, M.H.; Dey, A.R. An investigation about the possible role of cattle and goats as reservoir hosts for *Leishmania donovani* in Bangladesh. *J Vector Borne Dis* **2018**, *55*, 242–244. Doi: 10.4103/0972-9062.249484.
61. Paixão-Marques, M.D.S.; Alves-Martin, M.F.; Guiraldi, L.M.; Dos Santos, W.J.; de Lemos, F.A.; Sánchez, G.P.; Richini-Pereira, V.B.; Lucheis, S.B. First isolation of *Leishmania infantum* by blood culture in bovines from endemic area for canine visceral leishmaniasis. *Parasitology* **2019**, 1–3. Doi: 10.1017/S0031182019000088.
62. Lamattina, D.; Nava, S. Ticks infesting humans in Northern Misiones, Argentina. *Medicina (B Aires)* **2016**, *76*, 89–92.
63. Walker, A.R.; Bouattour, A.; Camicas, J.-L.; Estrada-Pena, A.; Horak, I.G.; Latif, A.A.; Pegram, R.G.; Preston, P.M. *Ticks of domestic animals in Africa: a guide to identification of species*; illustrée.; Bioscience Reports: Wisconsin University- Madison, **2003**; ISBN 0-9545173-0-X.
64. Nielsen, E.M.; Andersen, M.T. Detection and Characterization of Verocytotoxin-Producing *Escherichia coli* by Automated 5′ Nuclease PCR Assay. *J Clin Microbiol* **2003**, *41*, 2884–2893. Doi: 10.1128/JCM.41.7.2884-2893.2003.
65. Regnery, R.L.; Spruill, C.L.; Plikaytis, B.D. Genotypic identification of rickettsiae and estimation of intraspecies sequence divergence for portions of two rickettsial genes. *J. Bacteriol.* **1991**, *173*, 1576–1589.
66. Choi, Y.-J.; Lee, S.-H.; Park, K.-H.; Koh, Y.-S.; Lee, K.-H.; Baik, H.-S.; Choi, M.-S.; Kim, I.-S.; Jang, W.-J. Evaluation of PCR-Based Assay for Diagnosis of Spotted Fever Group Rickettsiosis in Human Serum Samples. *Clin Diagn Lab Immunol* **2005**, *12*, 759–763. Doi: 10.1128/CDLI.12.6.759-763.2005.
67. Hornok, S.; Földvári, G.; Elek, V.; Naranjo, V.; Farkas, R.; de la Fuente, J. Molecular identification of *Anaplasma marginale* and rickettsial endosymbionts in blood-sucking flies (Diptera: Tabanidae, Muscidae) and hard ticks (Acari: Ixodidae). *Vet. Parasitol.* **2008**, *154*, 354–359. Doi:

10.1016/j.vetpar.2008.03.019.

68. Loh, S.-M.; Gofton, A.W.; Lo, N.; Gillett, A.; Ryan, U.M.; Irwin, P.J.; Oskam, C.L. Novel *Borrelia* species detected in echidna ticks, *Bothriocroton concolor*, in Australia. *Parasit Vectors* **2016**, *9*. Doi: 10.1186/s13071-016-1627-x.
69. Albuquerque, A.; Campino, L.; Cardoso, L.; Cortes, S. Evaluation of four molecular methods to detect *Leishmania* infection in dogs. *Parasit Vectors* **2017**, *10*, 57. Doi: 10.1186/s13071-017-2002-2.
70. Thompson, J.D.; Higgins, D.G.; Gibson, T.J. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **1994**, *22*, 4673–4680.
71. Tamura, K.; Nei, M. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Mol. Biol. Evol.* **1993**, *10*, 512–526. Doi: 10.1093/oxfordjournals.molbev.a040023.
72. Kumar, S.; Stecher, G.; Tamura, K. MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Mol. Biol. Evol.* **2016**, *33*, 1870–1874. Doi: 10.1093/molbev/msw054.
73. Mixson, T.R.; Campbell, S.R.; Gill, J.S.; Ginsberg, H.S.; Reichard, M.V.; Schulze, T.L.; Dasch, G.A. Prevalence of *Ehrlichia*, *Borrelia*, and Rickettsial Agents in *Amblyomma americanum* (Acari: Ixodidae) Collected from Nine States. *Journal of Medical Entomology* **2006**, *43*, 1261–1268. Doi: 10.1603/0022-2585(2006)43[1261:POEBAR]2.0.CO;2.
74. Trout Fryxell, R.T.; DeBruyn, J.M. The Microbiome of *Ehrlichia*-Infected and Uninfected Lone Star Ticks (*Amblyomma americanum*). *PLoS One* **2016**, *11*. Doi: 10.1371/journal.pone.0146651.
75. Aktas, M.; Özübek, S. Transstadial Transmission of *Hepatozoon canis* by *Rhipicephalus sanguineus* (Acari: Ixodidae) in Field Conditions. *J. Med. Entomol.* **2017**, *54*, 1044–1048. Doi: 10.1093/jme/tjx050.
76. Hamšíková, Z.; Silaghi, C.; Rudolf, I.; Venclíková, K.; Mahríková, L.; Slovák, M.; Mendel, J.; Blažejová, H.; Berthová, L.; Kocianová, E.; et al. Molecular detection and phylogenetic analysis of *Hepatozoon* spp. in questing *Ixodes ricinus* ticks and rodents from Slovakia and Czech Republic. *Parasitol Res* **2016**, *115*, 3897–3904. Doi: 10.1007/s00436-016-5156-5.
77. Hailemariam, Z.; Krücken, J.; Baumann, M.; Ahmed, J.S.; Clausen, P.-H.; Nijhof, A.M. Molecular detection of tick-borne pathogens in cattle from Southwestern Ethiopia. *PLoS One* **2017**, *12*. Doi: 10.1371/journal.pone.0188248.
78. Laird, J.S.; Kocan, A.A.; Kocan, K.M.; Presley, S.M.; Hair, J.A. Susceptibility of *Amblyomma americanum* to natural and experimental infections with *Theileria cervi*. *J. Wildl. Dis.* **1988**, *24*, 679–683. Doi: 10.7589/0090-3558-24.4.679.
79. Mans, B.J.; Pienaar, R.; Latif, A.A. A review of *Theileria* diagnostics and epidemiology. *International Journal for Parasitology: Parasites and Wildlife* **2015**, *4*, 104–118. Doi: 10.1016/j.ijppaw.2014.12.006.
80. Njiiri, N.E.; Bronsvoort, B.M. deC; Collins, N.E.; Steyn, H.C.; Troskie, M.; Vorster, I.; Thumbi, S.M.; Sibeko, K.P.; Jennings, A.; van Wyk, I.C.; et al. The epidemiology of tick-borne haemoparasites as determined by the reverse line blot hybridization assay in an intensively studied cohort of calves in western Kenya. *Vet. Parasitol.* **2015**, *210*, 69–76. Doi: 10.1016/j.vetpar.2015.02.020.
81. Oura, C. a. L.; Bishop, R.P.; Wampande, E.M.; Lubega, G.W.; Tait, A. Application of a reverse line blot assay to the study of haemoparasites in cattle in Uganda. *Int. J. Parasitol.* **2004**, *34*, 603–613. Doi: 10.1016/j.ijpara.2003.12.012.



964